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Pharmaceutical quality of antimalarial drugs: quinine sulfate and Artemether/Lumefantrine tablets sold on Bukavu Market

Aladin Ombeni Mahano¹, Aline Zawadi Mahano¹, Nelson Hendwa Cubaka¹, Félicien Mushagalusa Kasali¹, Benjamin Bavurhe Zirirane¹, Lucien Murhula Namegabe¹, Pacifique Murhula Hamuli¹ and Ntokamunda Justin Kadima^{1,2*}

Abstract

Background: Generic antimalarial drugs sold in sub-Saharan Africa require tighter control as counterfeiting has grown more and more out of control. The study aimed to analyze the pharmaceutical quality of quinine sulfate (QS) and Artemether/Lumefantrine(AL) tablets marketed in Bukavu city compared to the current trends in other African cities.

Results: The samples were purchased in community pharmacies or from ambulatory street vendors and analyzed using a set of thirteen simple tests, including visual inspection, UV spectrometry, TLC, and conventional quality control procedures. More than 93% of AL samples had an acceptable global quality score of > 90%. Around 16.6% of QS batches did not satisfy the requirements about hardness, friability, and mass uniformity. Only 33.3% met the disintegration quality; 33.3% did not contain quinine; 8.33% had an active ingredient other than quinine.

Conclusion: The findings strongly alert the circulation of fake antimalarial medicines observed in many countries. Simple TLC procedures may help to detect any low-quality generics to avoid microbial resistance and guarantee the health of the population. Pharmacists and regulatory authorities are alerted to the circulation of low-quality generic quinine preparations in the country.

Keywords: Quinine, Artemether, Lumefantrine, Quality control, Congo

Background

Around 3.2 billion people, or almost half of the world's population, are exposed to malaria risk [1–4]. According to the World Malaria Report (WMR), an estimated 228 million malaria cases occurred worldwide in 2018, and 19 countries in sub-Saharan Africa and India carried almost 85% of the global malaria burden [1]. Of an estimated 405,000 deaths from malaria globally, nearly 85%

live in 20 countries in the WHO African Region, mainly Nigeria (24%) and the Democratic Republic of Congo (DRC) (11%) [5, 6]. Children aged under 5 years are the most vulnerable group affected by malaria; they accounted for 67% (272,000) of all malaria deaths worldwide in 2018.

Access to adequate and quality antimalarial drugs (AMDs) plays a crucial role in efforts to ensure complete cures and avoid vicious complications [7]. Without treatment, malaria can quickly lead to death from the circulatory disorders it causes. Also, in many parts of the world, malaria parasites have become resistant to several

* Correspondence: kadima48@yahoo.com; ntokamunda13@gmail.com

¹Department of Pharmacy, Faculty of Pharmaceutical Sciences and Public Health, Official University of Bukavu, Bukavu, Democratic Republic of Congo

²Department of Pharmacy, School of Medicine and Health Sciences, University of Rwanda, Kigali, Rwanda

AMDs, due in significant part to fake medications [8–10]. Among those counterfeit medicines, some contain sub-therapeutic doses and others not at all [11]. According to the molecular quantitative similarity analysis (MQSA) study, 25–50% of AMDs are of low quality, adulterated, or inferior quality. The figures from the US Food and Drug Administration (USFDA) indicate that up to 25% of all drugs consumed in emerging countries are counterfeit or substandard [12–14]. Counterfeiting affects all classes of drugs, but mainly antibiotics and AMDs, for which the demand is high in African countries where malaria is endemic. That demand leads to an informal trade in counterfeit products beyond the control of health authorities.

In the DRC, an estimated 97% of the population lives in zones with stable malaria transmission lasting 8 to 12 months per year; the other 3% is exposed to epidemic malaria in the high mountains of the east of the country [15, 16]. Around 80% of deaths occur at home, which represents millions of cases of death per year. The prevalence of fever in children under 5 years of age is 42%, corresponding to between 6 and 10 malaria episodes per child per year. The combination Artemether/Lumefantrine (CoArtem) and Artesunate/Amodiaquine (AS-AQ) are the most widely used AMDs in the initial treatment of non-severe *Falciparum* malaria and quinine for severe cases [17]. The circulation of counterfeit or poor-quality products is beyond control. This study aimed to assess the quality of generic quinine sulfate (QS) and Artemether-Lumefantrine (AL) tablets marketed in Bukavu using simple usual analytical techniques.

Methods

Study design

The study took place in Bukavu city, the capital of South-Kivu province in DRC, from May to September 2019. It assessed pharmaceutical technology requirements concerning labeling, physical examination, disintegration time, friability, and mass uniformity. Thin-layer chromatography (TLC), non-aqueous protometry, and UV-spectrophotometry served to identify and assay active ingredients using referral pharmacopeial guidelines and regulations. Critical attributes were labeling quality, general tests on pharmaceutical dosage forms, content and dissolution, identification, and the presence of unidentifiable impurities in formulations.

Equipment, chemicals, and reagents

The equipment included UV spectrophotometer (Pharmacia LKB Ultraspec Plus, Germany), chromatographic plates (pre-coated TLC sheets Alugram® Xtra Sil G/UV 254 layer: 0.20 mm silica gel 60 with fluorescent indicator UV 254), UV lamp 254 nm (Technology Transfer, Germany), analytical balance (balance digital jewelry

scale, capacity 20 g, readability 0.001 g, China), friability device (manufactured by lab-line instruments. Inc. Model n°1641 China), aggregation device (Erweka-apparate bau, Germany), durometer (Schleuniger-2E, Switzerland), hot plate (Protherm Winn Leek, Holland), water bath (Gerhardt Bonn, Germany), rotavapor (Pleuger, Switzerland), centrifuge (IEC Centra-2 centrifuge, USA), and vortex shaker (Vortex-2 genie Bohemia, USA).

The solvents were distilled water; toluene R (Merck KGaA, 64271 Darmstadt, Germany), ethyl acetate R (Merck KGaA, 64271 Darmstadt, Germany), methanol R (Maprochim-Labo, 59, lot 0112, 90% 0.791, DRC), hydrochloric acid R (PANREAC Quimica SAU, E-08211, Spain), glacial acetic acid R (BN: 04H240011, Spain), acetic anhydride R (E. Merck Darmstadt, Germany), acetone R (Merck KGaA 64271 Darmstadt, Germany), perchloric acid R (70%, density 1.76, E. Merck, D-6100 Darmstadt, Germany), chloroform R (Prolabo, Belgium), potassium iodide R (Merck, D-6100 Darmstadt, Germany), and mercury chloride II R (B Zedelgem, Germany).

Method development

Uniformity of mass

It represents the average mass of 20 tablets. The deviation of the individual tablet weight from the mean weight should not exceed a minimum of $\pm 5.0\%$ and a maximum of $\pm 10.0\%$ for 18 and 20 tablets, respectively [18].

Friability of tablets

A device manufactured by Lab-Line Instruments Inc. Model n°1641 operating by rotation at 100 revolutions per 4 min served to measure friability, with a total of tablets corresponding, as near as possible, to 6.5 g introduced in the machine. According to the requirement, the maximum acceptable loss of mass (obtained from a single test or the mean of 3 tests) is not greater than 1.0% for the sample [19].

Hardness-resistance of tablets to crushing

A durometer (Schleuniger-2E, Switzerland) served to measure the minimum and maximum force (newton) to crush each of 10 tablets [20].

Disintegration of tablets

We used a 6-tube basket disintegrating device Erweka-apparate, containing HCl 1 N as immersion fluid maintained at $37 \pm 2^\circ\text{C}$, in triplicate. The basket filled with six tablets was rotated for 30 min and then raised to count the units not completely disintegrated. At least 16 of the 18 dosage units tested should disintegrate in time [21, 22].

Identification of quinine sulfate by Mayer reagent

A tiny amount of crushed tablets powder was mixed with distilled water, filtered, and the filtrate mixed with the Mayer reagent (5 g of potassium iodide (KI), 1.36 g of mercury chloride (HgCl_2), and 100 ml of distilled water) in a test tube. The reaction was positive if a yellowish-white precipitate is formed [23].

Identification of quinine sulfate and Artemether-Lumefantrine by TLC

The protocol followed the existing procedures with slight adaptation. For QS samples [24–26], the stationary phase was silica gel R6; the mobile phase was methanol/ammonia (20: 0.5 v/v). The final concentration was 1.428 mg/ml. For AL samples [27, 28], the stationary phase was silica gel R6; the mobile phase was toluene/ethyl acetate/anhydrous acetic acid (18:4:2 v/v/v). The final concentration was 1.30 mg/ml of Artemether and 7.80 mg/ml of Lumefantrine. Approximately 2 μl of samples and 2 μl of standards were spotted at 1.5 cm on silica gel R6 plates. After migration, each chromatographic plate was removed, dried, and then examined with ultraviolet light (254 nm) for QS samples. The identification of AL samples required spraying the plate with sulfuric acid R/methanol R mixture (10:190 v/v) and then heating the chromatographic plate to dryness on the heating plate at 50 °C for approximately 10 min. We compared the correspondence in position, appearance, and intensity between the test samples and the standards. The retardation factor error (%Rf error) of Rf-sample and Rf-standard was calculated as follows:

$$\%R\text{Error} = \frac{100 \times (Rf_{\text{standard}} - Rf_{\text{sample}})}{Rf_{\text{standard}}}$$

Rule: If $Rf_{\text{error}} \leq 5\%$, the sample is considered valid; if Rf_{error} is between 5 and 10%, the analyte is deemed doubtful; if $Rf_{\text{error}} \geq 10\%$, it is invalid [2].

Quantitative analysis of quinine sulfate in tablets

Aliquots of 20 tablets crushed to a powder, equivalent to 100 mg of quinine sulfate, were gently stirred for 15 min in 40 ml of acetic anhydride R and 40 ml of anhydrous acetic acid. And we titrated with perchloric acid (0.1 mol/L) using violet crystal as the indicator (from violet to blue and apple green). Each milliliter of perchloric acid (0.1 mol/l) is equivalent to 26.10 mg of quinine sulfate [$(\text{C}_{20}\text{H}_{24}\text{N}_2\text{O}_2)_2, \text{H}_2\text{SO}_4 \cdot 2\text{H}_2\text{O}$]. The test requires that the yield be not less than 90.0% and not more than 110.0% of the amount of quinine sulfate [29, 30].

Quantitative analysis of Artemether-Lumefantrine

The principle consists of extracting Artemether and Lumefantrine from TLC spots and measuring the

absorbance at 254 nm. For the Artemether (sample and standard), the layer obtained by TLC was carefully scraped, weighed, and diluted with 2 ml of the HCl/ethanol mixture (1 mol/l) in a test tube. After homogenization on the vortex, the test tube was heated in a water bath at 55 °C for 5 h, then cooled to room temperature and centrifuged. The procedure was the same for the blank without Artemether. The resulting solution's absorbance was measured in a 1-cm layer quartz cell at 254 nm against a solvent cell containing the blank. The dilution was 38.5 for Artemether ($\text{C}_{16}\text{H}_{26}\text{O}_5$) in the sample and standard, using the absorptivity value $A_{1\text{cm}}^{1\%} = 385$ [27]. The percentage content of Artemether ($\text{C}_{16}\text{H}_{26}\text{O}_5$) in the tablets was calculated as follows:

$$\text{Yield}\% = \frac{100 \times (\text{mean Absorbance sample})}{\text{mean Absorbance standard}}$$

The test met the requirement if the yield is between 95.0 and 105.0% of the amount of Artemether ($\text{C}_{16}\text{H}_{26}\text{O}_5$) and Lumefantrine ($\text{C}_{30}\text{H}_{32}\text{Cl}_3\text{NO}$). For the lumefantrine analysis, the TLC layer spots were carefully scraped, weighed, and dissolved in 2 ml of methanol R in a test tube. The mixture was then homogenized on the vortex for 15 min and centrifuged. The absorbance was measured at 380 nm [28] against the blank prepared in the same way without Lumefantrine.

Data analysis

We set the score of the critical quality attributes as 1 or 0 when the test met or not the requirement—the sum of scores allowed calculating the percentages of global quality satisfaction. The quality was excellent (0.90–1.00), good (0.80–0.90), acceptable (0.70–0.80), and low (0.30–0.70), based on a modified psycho-physical Harrington's scale of quality [31].

The risk of treatment failure, death due to untreated disease, or toxicity was high, moderate, or low concerning the absence or wrong active ingredient, identity, under-dose, over-dose, disintegration, uniformity of mass, and labeling outcomes. MS Excel 2013 was used to calculate the descriptive statistics.

Results**Labeling of medicines**

The samples of generic QS and AL tablets analyzed were from the community pharmacies and ambulatory street vendors. As shown in Table 1, QS samples consisted of 12 batches containing 30 tablets each from 5 brands. AL samples consisted of 12 packs containing 24 tablets per blister from two brands. As presented, the visual inspection shows that 4 (30%) batches of QS samples (QSB2.2, QSB2.4, QSB3.5, and QSB4.6) did not meet all the

Table 1 Samples of quinine sulfate and Artemether-Lumefantrine brands tested

Quinine sulfate tablets samples					Artemether-Lumefantrine tablets samples				
N	Batch	Dose, mg	Shelf life	Country	N	Batch	Dose, mg	Shelf life	Country
1	QSB1.1	500	5 years	Democratic Republic of Congo	1	ALB1.1	20/120	3 years	Uganda
2	QSB1.3	500	5 years	Democratic Republic of Congo	2	ALB1.4	20/120	3 years	Uganda
3	QSB1.8	500	5 years	Democratic Republic of Congo	3	ALB1.5	20/120	3 years	Uganda
4	QSB1.9	500	5 years	Democratic Republic of Congo	4	ALB1.7	20/120	3 years	Uganda
5	QSB1.11	500	5 years	Democratic Republic of Congo	5	ALB1.8	20/120	3 years	Uganda
6	QSB2.2	500	6 years	Missing	6	ALB1.9	20/120	3 years	Uganda
7	QSB2.4	500	2 years	Missing	7	ALB1.11	20/120	3 years	Uganda
8	QSB3.5	500	Missing	Missing	8	ALB1.12	20/120	3 years	Uganda
9	QSB4.6	500	Missing	Missing	9	ALB2.2	20/120	3 years	Turkey
10	QSB5.7	500	5 years	Netherlands	10	ALB2.3	20/120	3 years	Turkey
11	QSB5.10	500	4 years	Netherlands	11	ALB2.6	20/120	3 years	Turkey
12	QSB5.12	500	5 years	Netherlands	12	ALB2.10	20/120	3 years	Turkey

labeling requirements. Some information was inaccurate or missing. The labeling on the packages of AL samples contained all the needed information.

Manufacturing technology quality

The elements tested were mass uniformity, hardness, friability, and disintegration time. For QS samples, two batches (QSB4.6, QSB5.7) did not meet the mass uniformity criteria because more than three tablets deviated beyond $\pm 10.0\%$. The hardness of 2 batches (QSB1.3, QSB3.4) did not lay between 50 and 150 N. The disintegration time of 8 batches (QSB1.1, QSB1.3, QSB1.8, QSB1.9, QSB1.11, QSB2.2, QSB2.4, and QSB3.5) was out of specification limits. For AL samples, two batches (ALB1.4, ALB2.2) did not meet the mass uniformity criteria. All batches satisfied the specification limits of hardness, friability, and disintegration.

Identification of active ingredients

Figure 1 shows the typical TLC chromatograms obtained.

Identification of QS samples showed that four batches (QSB5.7, QSB5.10, QSB5.12, QSB4.6) did not contain quinine sulfate; QSB4.6 had an active ingredient other than quinine sulfate because the $R_{f_{error}}$ was $> 10\%$. Four batches of QS samples failed Mayer's reaction (QSB4.6, QSB5.7, QSB5.10, and QSB5.12).

Content assay of active ingredients

Table 2 shows the results of quantitative assays. For QS samples, the batches QSB4.6, QSB4.7, QSB4.10, and QSB4.12 did not contain quinine (yield = 0%). The qualitative analysis revealed the absence or insufficient amount of active ingredients or the presence of other components than quinine sulfate in some batches.

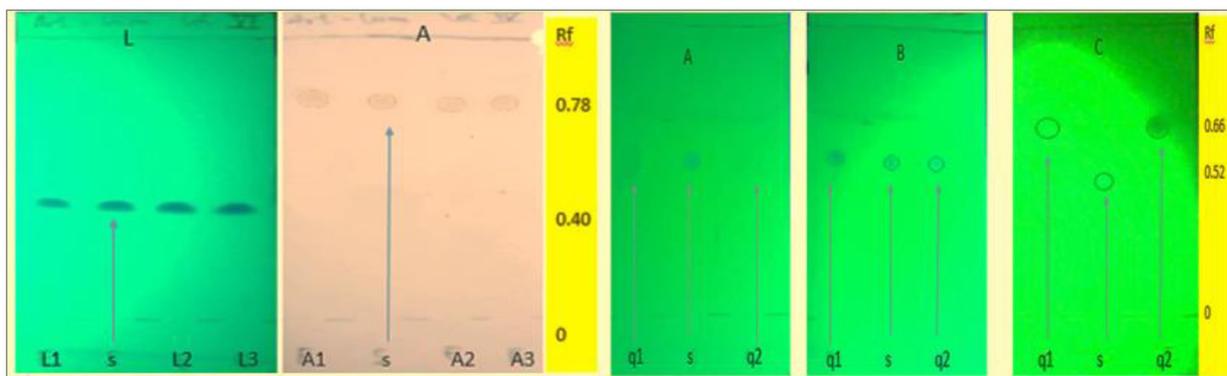


Fig. 1 Typical TLC chromatograms of Artemether (A)/Lumefantrine (L) samples at the left and three quinine samples (A, B, C) at right. The TLC of Lumefantrine (L) and Artemether (A) shows that all batches in triplicate spots gave R_f similar to the standards(s), with $R_{f_{error}} < 2\%$ for all the spots. However, the TLC of quinine shows the absence of quinine sulfate in sample spot A(q2); the similarity of B(q1) and B(q2) spots with the standard (s); the presence of other products different from the quinine standard in C(q1) and C(q2)

Table 2 Results of quantitative assays

QS batches	Quinine	AL batches	Artemether	Lumefantrine
Batches	Yield (%)	Batches	Yield (%)	Yield (%)
QSB1.1	96.51 ± 1.99	AL B1.1	99.46 ± 2.93	98.15 ± 0.49
QSB1.3	94.32 ± 2.61	AL B1.4	100.10 ± 1.76	98.89 ± 0.82
QSB1.8	101.31 ± 1.51	ALB1.5	100.19 ± 1.91	80.37 ± 0.40*
QSB1.9	93.45 ± 3.01	ALB1.7	91.81 ± 1.50	85.61 ± 0.47*
QSB1.11	98.69 ± 3.01	ALB1.8	90.06 ± 1.65	87.38 ± 1.23*
QSB2.2	94.32 ± 2.61	ALB1.9	93.99 ± 0.58	99.73 ± 0.97
QSB2.4	98.69 ± 3.01	ALB1.11	94.3 ± 0.29	96.13 ± 0.35
QSB3.5	99.46 ± 1.52	ALB1.12	97.05 ± 0.45	98.46 ± 0.98
QSB4.6	0*	ALB2.2	96.78 ± 0.33	100.37 ± 0.59
QSB5.7	0*	ALB2.3	100.44 ± 1.97	82.78 ± 0.56*
QSB5.10	0*	ALB2.6	97.90 ± 2.64	94.44 ± 0.62
QSB5.12	0*	ALB2.10	97.84 ± 2.40	98.70 ± 0.62

*Out-of-specifications

For AL samples, the yield of Lumefantrine batches ALB1.5, ALB1.7, ALB1.8, and ALB2.3 was < 90%.

Global quality scores

Figure 2 shows the satisfaction of each QS batch to the 14 tests performed, using score 1 (if the requirement is satisfied) or 0 (if not) to evaluate the critical quality attributes.

It appears that no quinine batch obtained a 100% score; the maximum score was 92.9%, corresponding to one failure (QSB1.11, QSB1.8, QS1.1), and the minimum was 35.7%, corresponding to 9 fails (QS4.6). For example, batches QSB2.2, QSB2.4, QSB3.5, and QSB4.6

(from brands 2, 3, and 4) did not satisfy all required labeling information; all batches of brands 1, 2, and 3 failed the disintegration test; all samples of brands 4 and 5 did not contain quinine. The most failed tests were disintegration (33.3%) and assay (41.7%).

Figure 3 shows the satisfaction of each AL batch to the 13 tests performed, also using score 1 (if the requirement is satisfied) or 0 (if not) to evaluate the critical quality attributes. All labeling information needed was given. However, only five batches satisfied the assay test (ALB1.1, ALB1.4, ALB1.12, ALB2.2, ALB2.10). The minimum score for AL batches was 92.3%. The most failed test was the assay (41.7%).

Discussion

Counterfeiter medicines are the leading cause of resistance, morbidity, mortality, and dysfunction failure of the public health care system interest [32–36]. Quinine and Artemether/Lumefantrine fixed-dose are on the WHO essential medicines model list for curative malaria [32]. In this study, we assessed the pharmaceutical quality of QS and AL tablets under the following critical quality requirements: labeling quality, general tests on pharmaceutical dosage forms, content and dissolution, identification, and the presence of unidentifiable impurities in formulations. The findings vigorously back the importance of the counterfeiting phenomenon as described in numerous studies.

Drug labeling is a legislative requirement to obtain information on drugs such as brand name, pharmaceutical form, manufacture and expiration dates, administration route, manufacturer, and country manufacturing [37].

Batch	DOSE	NO BATCH	ISSUE	EXPIRE	SHELF	COMPANY	COUNTRY	UNIFORMITY	HARDNESS	FRIABILITY	DISINTEGRATION	TLC	ASSAY	MAYER	TOTAL	%
QSB1.1	1	1	1	1	1	1	1	1	1	1	0	1	1	1	13	92.9
QSB1.3	1	1	1	1	1	1	1	1	0	0	0	1	0	1	10	71.4
QSB1.8	1	1	1	1	1	1	1	1	1	1	0	1	1	1	13	92.9
QSB1.9	1	1	1	1	1	1	1	1	1	1	0	1	0	1	12	85.7
QSB1.11	1	1	1	1	1	1	1	1	1	1	0	1	1	1	13	92.9
QSB2.2	1	1	1	1	1	0	0	1	1	0	0	1	0	1	9	64.3
QSB2.4	1	1	1	1	1	0	0	1	0	0	0	1	1	1	9	64.3
QSB3.5	1	0	0	1	0	0	0	1	1	1	0	1	1	1	8	57.1
QSB4.6	1	0	0	1	0	0	0	0	1	1	1	0	0	0	5	35.7
QSB5.7	1	1	1	1	1	1	1	0	1	0	1	0	0	0	9	64.3
QSB5.10	1	1	1	1	1	1	1	1	1	1	1	0	0	0	11	78.6
QSB5.12	1	1	1	1	1	1	1	1	1	1	1	0	0	0	11	78.6
Total	12	10	10	12	10	8	8	10	10	8	4	8	5	8		
%	100	83.3	83.3	100	83.3	66.7	66.7	83.3	83.3	66.7	33.3	66.7	41.7	66.7		

Fig. 2 Quality compliance scores of quinine batches analyzed. Dose (the content strength), no batch (batch number), issue (the issued date), expire (the expiring date), shelf (the shelf life), company (the manufacturer), country (the country of production), uniformity (the uniformity of mass), hardness (the resistance), friability (friction), disintegration (disintegration time), TLC (chromatogram), assay (dosage), and Mayer (identification of alkaloids)

Batch	DOSE	NO BATCH	ISSUE	EXPIRE	SHELF	COMPANY	COUNTRY	UNIFORMITY	HARDNESS	FRIABILITY	DISINTEGRATION	TLC	ASSAY	Total	TOTAL
ALB1.1	1	1	1	1	1	1	1	1	1	1	1	1	1	13	100
ALB1.4	1	1	1	1	1	1	1	0	1	1	1	1	1	12	92.3
ALB1.5	1	1	1	1	1	1	1	1	1	1	1	1	0	12	92.3
ALB1.7	1	1	1	1	1	1	1	1	1	1	1	1	0	12	92.3
ALB1.8	1	1	1	1	1	1	1	1	1	1	1	1	0	12	92.3
ALB1.9	1	1	1	1	1	1	1	1	1	1	1	1	0	12	92.3
ALB1.11	1	1	1	1	1	1	1	1	1	1	1	1	0	12	92.3
ALB1.12	1	1	1	1	1	1	1	1	1	1	1	1	1	13	100
ALB2.2	1	1	1	1	1	1	1	1	1	1	1	1	1	12	100
ALB2.3	1	1	1	1	1	1	1	1	1	1	1	1	0	12	92.3
ALB2.6	1	1	1	1	1	1	1	1	1	1	1	1	0	12	92.3
ALB2.10	1	1	1	1	1	1	1	1	1	1	1	1	1	13	100
Total	12	12	12	12	12	12	12	10	12	12	12	12	5		
%	100	100	100	100	100	100	100	83.3	100	100	100	100	41.7		

Fig. 3 Quality compliance scores of Artemether/Lumefantrine batches analyzed

Thus, the investigated products might not have a risk of confusion associated with labeling. The visual inspection results on the physical characteristics of tablets and labeling information revealed that QS products did not comply with the WHO guideline on the labeling for pharmaceutical products. All QS samples mentioned the dose; 83.3% mentioned batch number and date of issue and shelf life; only 66.7% mentioned the manufacturer and country. AL samples complied satisfactorily. A tighter inspection of the label can help suspect wrong products. Often, counterfeiters copy the brand's design or the designation of a product and sometimes the address. In developed countries, anticounterfeit packaging technologies are being developed [38–40]. The European trend is towards 2D barcodes (anticounterfeit technologies), and the USA has implemented radio frequency identification (RFID) [41]. The probability of detecting the failure in DRC is low.

According to Good Manufacturing Practice (GMP), uniform mass and friability are also critical attributes of quality [36, 42, 43]. Failure to quickly release the active ingredient for conventional formulations has the risk of therapeutic loss or delayed effect onset, explaining the number of deaths. The analysis of AL brands indicated that all of the samples but one complied with the quality specifications. However, for QS samples, only 3 out of 12 batches fully satisfied the critical technology attributes. Also, mass uniformity is an alternative test for content uniformity of uncoated and film-coated tablets. Here, defaults of two QS batches and one AL batch might not affect the content uniformity of the investigated products' dose units. Many studies show that most

antimalarial solid dosage forms pass the basic tests, such as the uniformity of mass for tablets, the content test, but a few pass in vitro product dissolution test required [36, 42, 43].

TLC and Mayer's QS identification revealed that 7 out of 12 batches did not contain the active principle. The results of quinine amount indicated that only 41.7% of investigated products did comply with the pharmacopoeial specification limit (90–110%). Contrarily, AL samples showed excellent scores (>90%) for Artemether, even though the content of Lumefantrine was under-dose in 4 batches. The risk of treatment failure due to bad quality is significantly higher with QS tablets than AL fixed-dose products. Studies carried out in Rwanda and the east of DR Congo had also shown the under-dosing and the absence of the quinine [11, 44]. One study conducted in DRC [45] on 150 AL samples collected from private pharmaceutical outlets in 8 main cities (Goma, Kikwit, Kinshasa, Kisangani, Lubumbashi, Matadi, Mbandaka, and Mbuji-Mayi) found 3 (2%) visual inspection failure and 4 (2.7%) TLC test failure. HPLC assays showed 46 (30.7%) samples had Artemether contents below 90% and 17 (11.3%) above 110% of the content claimed on the label; 32 (21.7%) samples had lumefantrine content below 90% and 8 (5.3%) above 110%. Studies in Kenya [46], Burkina Faso [47], and Ethiopia [48] also detected under-dose artemisinin generic products.

According to the WHO, counterfeit AMDs are often sold at low prices because they do not contain the right ingredients. Oral administration is the most widely accepted route due to its convenience in self-

administration and easy manufacturing, which lead to easy counterfeiting. In DRC, pharmacies and free markets are among the potential sellers of inexpensive anti-malarial to health facilities and the public. The unemployment rate is high; the average monthly household income remains low [49]. Such poverty leads people to look for cheaper antimalarial products sold in free markets. The situation makes it very hard to detect fake products quickly. Non-pharmacists own most pharmacies, and many are not at all managed by pharmacists. One knows that non-pharmacist health professionals are not qualified to detect the quality characteristics of the drugs they prescribe or administer.

Conclusion

The findings show that antimalarial drugs sold in the DRC pharmaceutical market are imported products from various countries, and not all brands are of good quality. African countries face multiple kinds of counterfeiting favored by poverty, lack of proxy control tools, and inefficacy of regulatory authority. That raises the importance of surveying the quality of all pharmaceutical products imported. Pharmacists should be involved at each step of the medication management system to detect any anomalies to minimize the counterfeiting phenomenon impact. Simple tests may be of significant help.

Abbreviations

AL: Artemether-Lumefantrine; ALB: Artemether-Lumefantrine fixed-dose batch; AMD: Antimalarial drug; DRC: Democratic Republic of the Congo; GMP: Good Manufacturing Practice; MQSA: Molecular quantitative similarity analysis; QS: Quinine sulfate; QSB: Quinine sulfate batch; RFID: Radio-frequency identification; TLC: Thin-layer chromatography; WHO: World Health Organization; WMR: World Malaria Report

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Authors' contributions

AOM and AZM designed the study and literature search. AZM, LMN, and PMH operated the pharmaceuticals testing. NHC, FMK, and BBZ conducted the identification tests. AOM and NJK revised the protocols and supervised and wrote the final draft. All authors have read and approved the manuscript.

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Ethics approval and consent to participate

Not applicable

Consent for publication

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Competing interests

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