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Virtual screening, pharmacokinetics, and molecular dynamics simulations studies to identify potent approved drugs for Chlamydia trachomatis treatment

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Abstract

Background: The most frequent bacterial sexually transmitted disease is Chlamydia trachomatis (STD). In 2010, the Centers for Disease Control and Prevention (CDC) received 1.3 million reports of cases (CDC). Human chlamydial infections are linked to a variety of clinical symptoms. Inclusion (IncA) membranes are a promising drug target for the treatment of Chlamydia trachomatis. In the present study, molecular docking, ADMET, golden triangle, and molecular dynamics (MD) simulation studies were performed on a series of salicylidene acylhydrazides derivatives against Chlamydia trachomatis. Three types of docking software with different algorithms were used to screen the potential candidate against Chlamydia trachomatis.

Results: The results obtained from the docking analysis succeeded in screening nine novel hit compounds with high affinity to IncA membranes. Then, pharmacokinetics properties were calculated to spot out the drug-likeness of the selected compounds. Also, golden triangles were performed on the selected compounds. Compounds outside the golden triangle indicate that they would have clearance problems. Out of the nine novel hits drugs, four compounds pass the golden triangle screening and virtually all the quality assurance tests proposed by the model and were used for further analysis. One-ns molecular dynamics simulations on the docked complex of compound 44 (one of the highly active selected compounds of the dataset) aided in the further exploration of the binding interactions. Some crucial residues such as Ser111, Gln114, Asn107, Leu142, Gly144, Gln143, Lys104, Tyr149, Phe108, Phe145, and Arg146 were identified. Conventional and carbon-hydrogen bond interactions with amino residues Arg146, Asn107, Phe145, and Ser111 were critical for the binding of inclusion (IncA) membranes inhibitors.

Conclusion: Outcomes of the study can further be exploited to develop potent inclusion (IncA) membranes inhibitors.

Keywords: Molecular docking, iGemDock, MVD, AutoDock Vina, ADMET, Golden triangle, Chlamydia trachomatis, Salicylidene acylhydrazides derivatives, MDs simulations

Background

Chlamydia trachomatis is the world's most prevalent cause of bacterial sexually transmitted infection and infectious blindness in endemic locations, primarily in Africa and the Middle East infecting approximately a hundred million people each year [1, 2]. Chlamydia trachomatis is a parasite that lives only inside cells. It goes

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through a replicative developmental growth cycle inside the cells of its host, which leads to cell death [3]. According to the World Health Organization, 92 million new instances of *Chlamydia trachomatis* infection occur each year [4]. In the USA, it is estimated that over 3 million chlamydial infections occur each year among sexually active adolescents and adults [5, 6]. From 1993 to 2011, the number of chlamydial infections reported to the Centers for Disease Control and Prevention (CDC) increased from 178 to 453.4 cases per 100,000 people [7]. Human illnesses caused by *Chlamydia trachomatis* include cervicitis, salpingitis, acute urethral syndrome, endometritis, ectopic pregnancy, infertility, and pelvic inflammatory disease (PID) in women; conjunctivitis and pneumonia in neonates; and urethritis, proctitis, and epididymitis in males [8]. Traditional and orthodox treatment of some existing drugs is ineffective and unable to stop the process of a fast-growing parasitic organelle inside the host cell called "Inclusion," which is poorly understood [9, 10]. The development of novel potential therapies for the treatment of *Chlamydia trachomatis* represents an important means of extending life span and improving quality of life [11, 12]. Antimicrobial resistance has been listed by the World Health Organization (WHO) as one of the three most serious threats to human health and a severe problem in many parts of the world [13]. *Chlamydia trachomatis* a Gram-negative bacterium stands as proof of high-level resistance to most categories of antibiotics [14]. Infections caused by multidrug-resistant *Chlamydia trachomatis* are on the rise in hospitals, especially in the intensive care unit (ICU), and are linked to greater expenses, increased morbidity, and high fatality rates [15]. Increased patient contact with health care personnel, increased severity of illness, overuse of existing antimicrobial agents, underlying conditions, exposure to multiple invasive devices and procedures, and crowding of patients in a small specialized area are all factors that contribute to an increased risk of infection in intensive care unit patients [16]. As a result, finding new strong, safer, and less expensive *Chlamydia trachomatis* or bacterial inhibitors has become the greatest problem the human race has ever faced. For this reason, the development of new drugs able to fight *Chlamydia trachomatis* is still in great interest.

Previous studies on anti-*Chlamydia trachomatis* compounds have been published; one is on the synthesis of substituted ring-fused 2-pyridines and evaluation for the ability to attenuate *Chlamydia trachomatis* infectivity [17]. In 2014, Abdelsayed and his colleagues design and synthesize a 3-isoxazolidone derivative and also evaluate the paths by which these new compounds affect *Chlamydia trachomatis* serovar L2 development in HeLa cells, in the presence or absence of exogenously added iron

[1]. Edache and his co-workers conducted quantitative structure–activity relationship (2D-QSAR), comparative molecular field analysis (CoMFA), and molecular dynamics simulations on novel thiazoline 2-pyridone amide derivatives [2]. In recent years, the use of molecular modeling has produced very impressive results in the drug discovery process [18].

In this place, we report the molecular modeling of salicylidene acylhydrazides derivatives to continue our ongoing research on *Chlamydia trachomatis* inhibitor molecular modeling. The docking simulation, molecular dynamics simulations, and ADMET filtering were applied to detect new potential candidates for *Chlamydia trachomatis* treatment.

Methods

In the present study, we have selected 58 salicylidene acylhydrazides derivatives from the PubChem database (www.ncbi.nlm.nih.gov/pubchem) with accession number AID 473049/473050 that have been already validated to have antimicrobial activity. All compounds were tested using the same experimental procedure, and a wide range of *Chlamydia trachomatis* inhibition activities were covered. Structures of the selected prevailing forms were optimized with the semiempirical PM3 method using the Spartan'14 software (www.wavefun.com) (Table 1). The optimized structures were saved for further analysis.

Virtual screening using multiple molecular docking tools

The strength of association or binding affinity between two molecules can be predicted by molecular docking techniques [19]. Molecular docking simulations were calculated from three different algorithms; therefore, it is assumed that the value represents a real condition. We chose 58 phytochemical substances that have been considered to have potential antibacterial activity for this study. The three-dimensional crystal structure of the IncA soluble domain was retrieved from RCSB (www.rcsb.org/pdb) protein data bank (PDB Code: 6e7e). The whole structure of the receptors was targeted for our molecular docking study except for heteroatoms that were detected from the receptors. Primary docking analysis was performed using iGemdock [20], which uses empirical scoring function and Generic Evolutionary Method for molecular docking. It comes with a graphical user interface that detects pharmacological interactions and does virtual screening. The following parameters for docking have been chosen: population size=200, number of generations=70, and number of solutions=3. To further increase the docking accuracy, by using a more complex scoring function, Molegro Virtual Docker (MVD) was used [21]. The imported structures must be properly prepared, that is, the atom connectivity and bond orders are correct, and partial atomic charges are assigned. PDB files often have the poor or missing assignment of explicit hydrogen, and the PDB file format cannot accommodate bond

Table 1 PubChem_CID number and corresponding activity pIC_{50} of salicylidene acylhydrazides derivatives

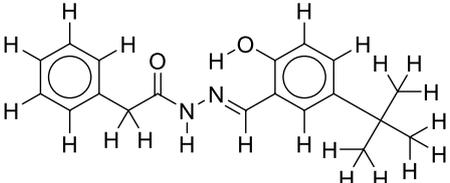
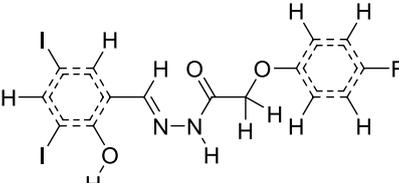
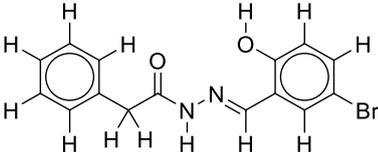
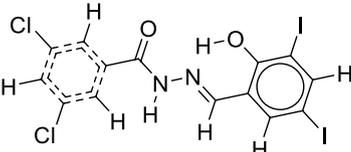
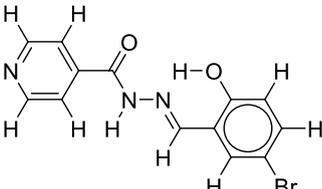
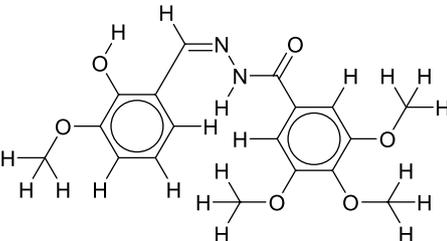
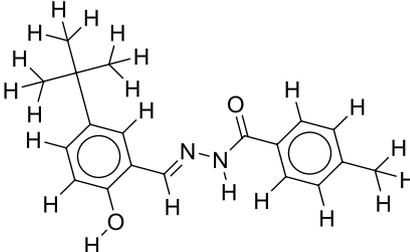
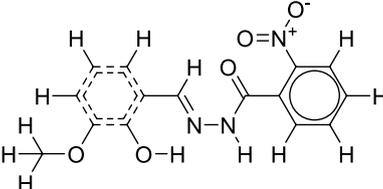
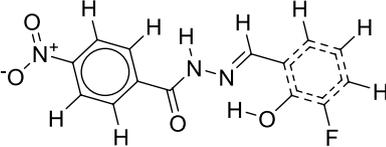
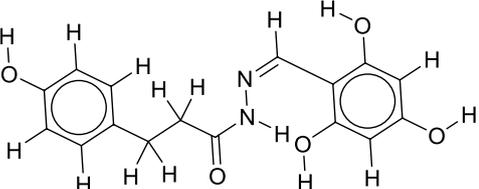
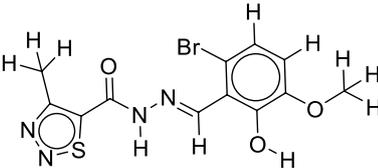
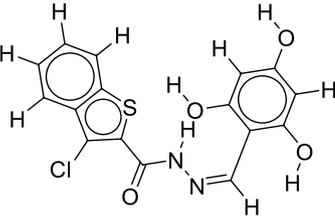
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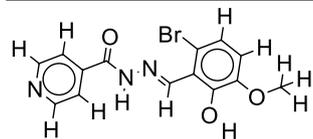
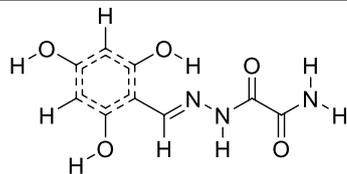
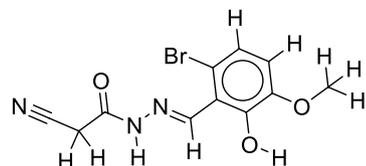
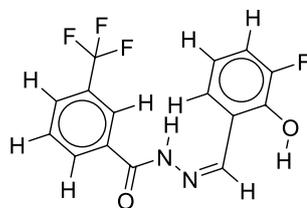
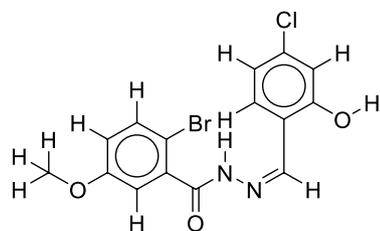
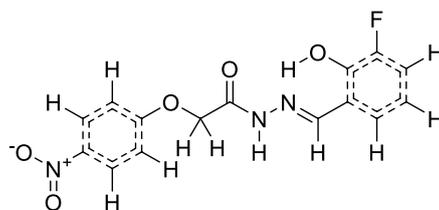
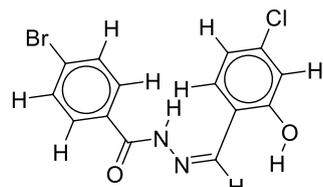
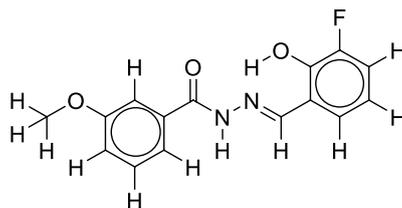
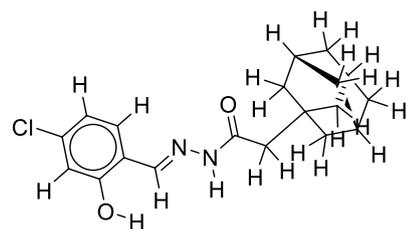
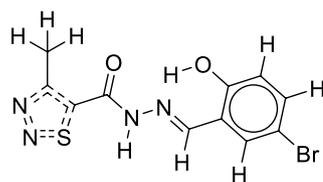
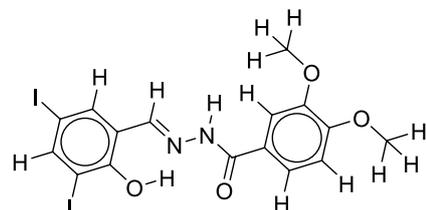
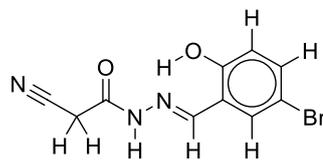
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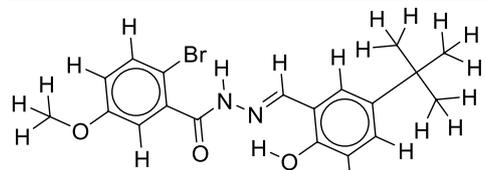
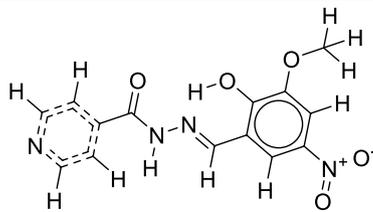
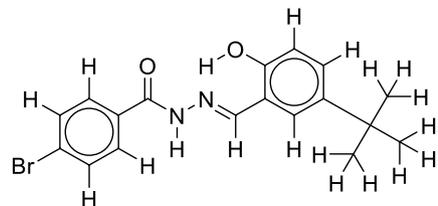
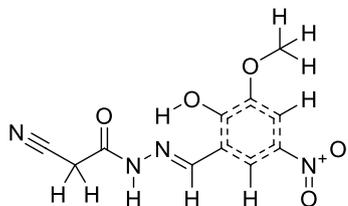
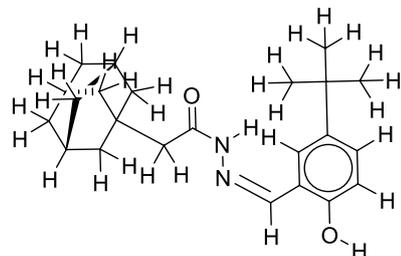
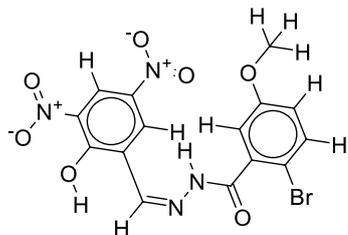
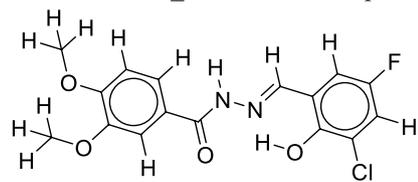
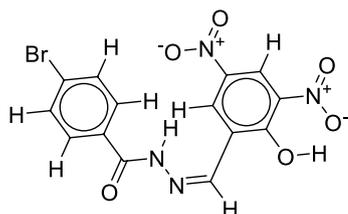
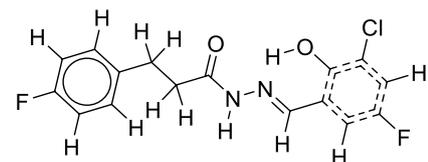
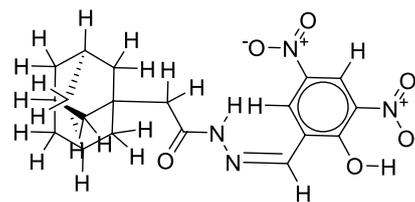
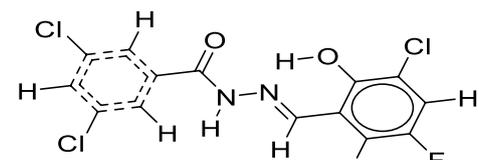
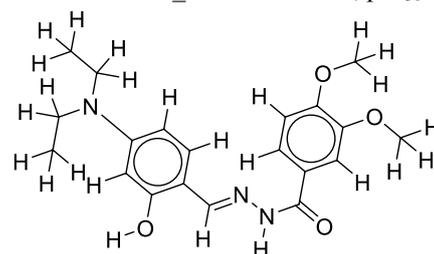
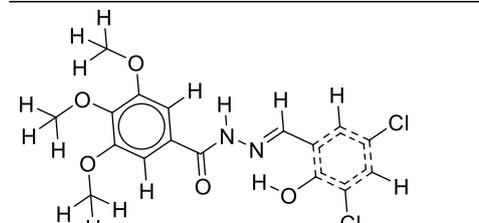
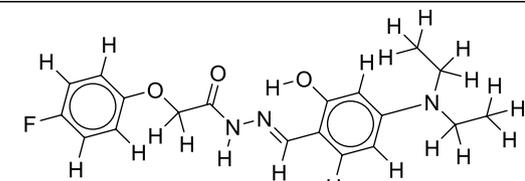
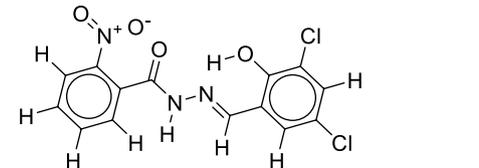
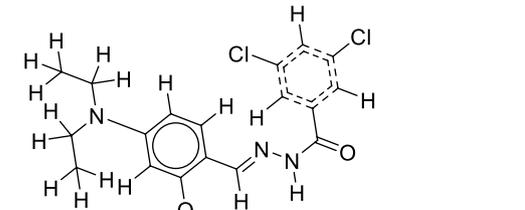
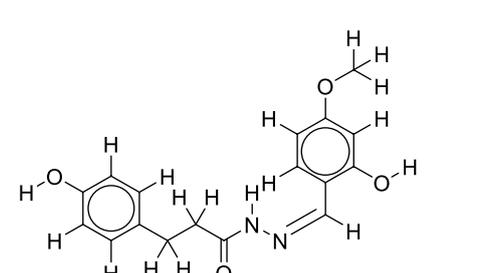
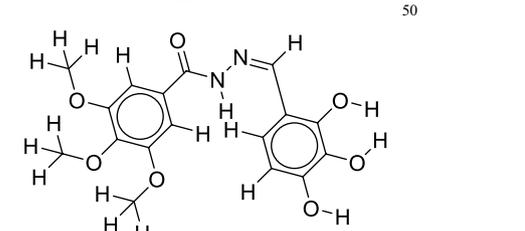
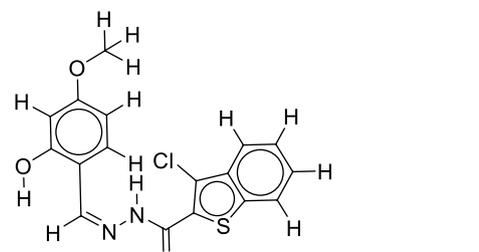
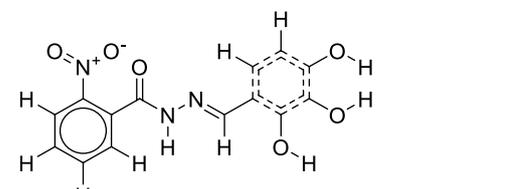
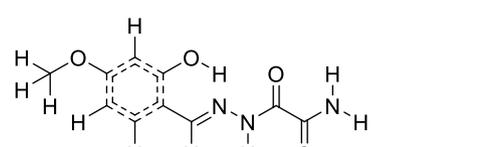
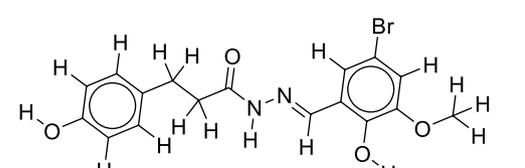
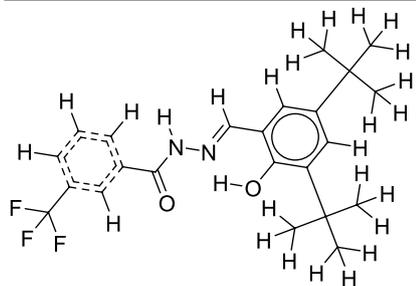
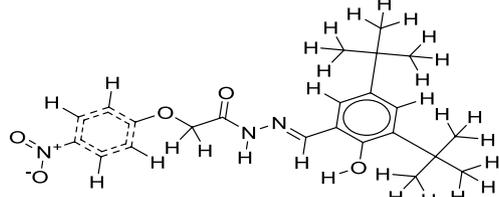
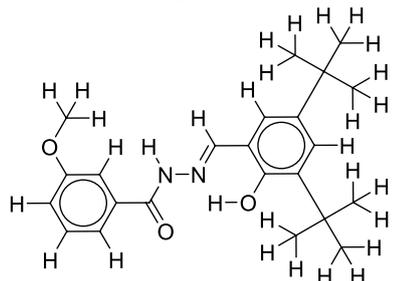
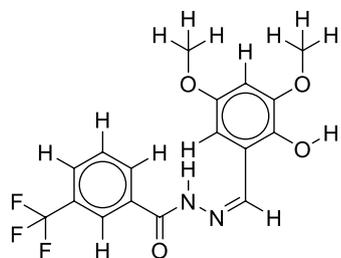
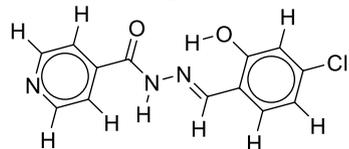
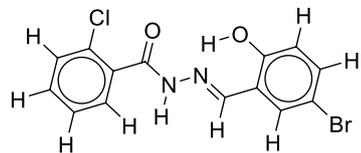
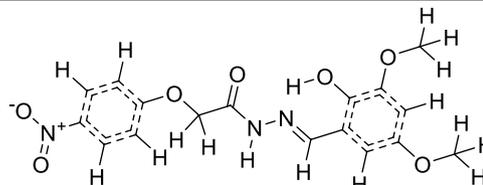
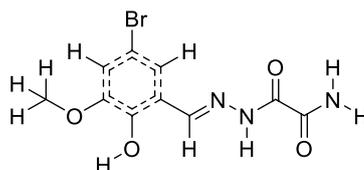
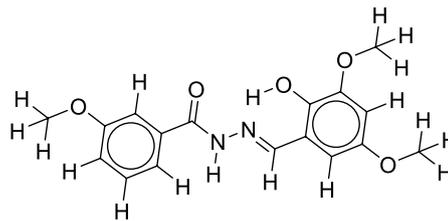
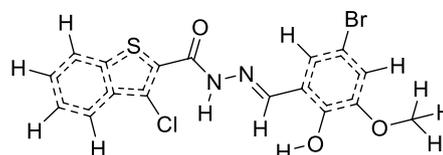
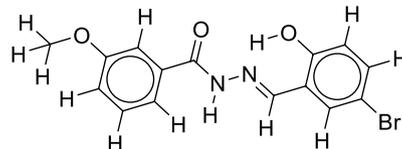
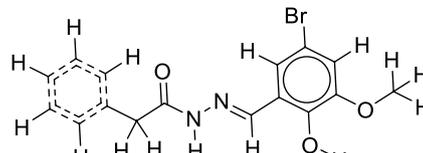
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 <p>35: PUBCHEM_CID 136167972; pIC₅₀=7.886</p>	 <p>50: PUBCHEM_CID 136167982; pIC₅₀=7.745</p>

order information. The utilities included in MVD were used to do all necessary valency tests and H-atom additions. The binding location is the area where the docking procedure will look for promising poses. The docking simulation results were then validated through AutoDock Vina [22] (PyRx software) with

the following setup: exhaustiveness=8; center_x=-2.3621; center_y=17.7247; center_z=36.6066; size_x=71.1413702774; size_y=32.6226404202; size_z=50.6705288601, and the interaction of all the docking approach was visualized through Discovery studio software.

Table 1 (continued)36: PUBCHEM_CID 136167973; pIC₅₀=8.22237: PUBCHEM_CID 136167974; pIC₅₀=7.92138: PUBCHEM_CID 136167975; pIC₅₀=7.85439: PUBCHEM_CID 136167976; pIC₅₀=7.40955: PUBCHEM_CID 136167986; pIC₅₀=7.56956: PUBCHEM_CID 135521406; pIC₅₀=7.95951: PUBCHEM_CID 136167983; pIC₅₀=7.74552: PUBCHEM_CID 135815951; pIC₅₀=8.00053: PUBCHEM_CID 136167984; pIC₅₀=7.69954: PUBCHEM_CID 136167985; pIC₅₀=7.63857: PUBCHEM_CID 135615174; pIC₅₀=7.65858: PUBCHEM_CID 135536260; pIC₅₀=8.046

ADMET, drug-likeness evaluation

Candidates for drugs should have positive ADME qualities and, ideally, be non-toxic. Using the DataWarrior software, the developed compounds were tested for their ADME profile, which included drug-likeness, partition coefficient, solubility, and several other metrics. In addition, the toxicity of the chosen drug was predicted.

Molecular dynamic (MD) simulation

The chosen complex was then simulated for 1 ns with VMD-NAMD software [23, 24]. To construct receptor molecular topology and configuration files, the Charmm-gui Web site was utilized. By adding 0.15 M KCl to the simulated system, the system was neutralized. TIP3P-BOX, a water molecules box, was then submerged in the neutralized system. Each system was equilibrated with NVT (constant number of particles, volume, and temperature) as well as NPT (constant number of particles, pressure, and temperature) before the production phase to guarantee an even distribution of solvent and ions surrounding the protein–ligand complex.

Results

Docking evaluations

Tables 2 and 3 reveal the numerical value of the total energy, van der Waals, hydrogen bond, and electrostatic of the standard drugs and the 58 salicylidene acylhydrazides derivatives with iGemDock v2.1 software. The docking interactions of the standard drugs and the best interacting ligand are reflected in Figs. 1 and 2.

With Molegro Virtual Docker (MVD) and PyRx software, all the standard drugs and ligands were docked with the protein structure and the best-docked molecules were listed depending on their MolDock score, Rerank score, hydrogen bond, and binding affinity as given in Tables 4 and 5. Figures 3 and 4 show detailed interaction of the standard drugs and the best four ligands with the MVD program, while Figs. 5 and 6 give in detail the docking pose of the reference or standard drugs and the four best ligands with PyRx software, respectively.

Table 2 The results of the reference drugs with iGemDock v2.1

SD	Total energy	VDW	HBond	Elec
Ampicillin	−92.5875	−64.7854	−27.8021	0
Oxytetracycline	−90.744	−62.8534	−27.8906	0
Chlortetracycline	−90.6709	−54.49	−36.1809	0
Ceftriaxone	−82.7819	−55.1418	−25.9849	−1.65517

SD, standard drugs

Pharmacokinetic evaluation

Lipinski's rule of 5 and Veber's rule of 2 of both the standard drugs and the selected compounds are reflected in Table 6. Table 7 shows the lipophilicity of the selected drugs, while Table 8 presents the toxicity risk factors of the standard drugs and the selected compounds.

Golden triangle and MD simulations

Figure 8A–C shows the variation in kinetic energy, total energy, potential energy versus time in 1-ns MD simulations, while Fig. 8D shows the binding interaction of the simulated complex.

Discussion

Docking screening of compounds with iGemdock software

Four anti-Chlamydia trachomatis drugs—ampicillin, oxytetracycline, chlortetracycline, and ceftriaxone, were used as a control for the following study. iGemdock calculated the total energy, van der Waals energy, electrostatic, and hydrogen bond of the four standard drugs when docking them into the enzymes. K-means and hierarchical clustering approaches are used by iGemdock's post-analysis tools. Table 2 shows the summary of docking results, and detailed interactions with the enzyme are shown in Fig. 1. Binding energies of the receptor–ligand interactions are very important to report how fit the drug binds to the target macromolecule. It can be calculated that according to iGemdock ampicillin is the best binding receptor because of its total energy of −92.5875 kcal/mol. A higher fitness score suggests better docking interaction between protein and ligand and will be acceptable of the compound as a drug.

The total energy of compounds 4, 38, 43, 44, and 48 was found to be −78.3402, −76.561, −78.9421, −87.3827, and −77.9572 kcal/mol, respectively (Table 3). From the results, a good number of standards (Table 2) displayed activity comparable to those selected compounds. Compound 44 has the best score and the most outstanding inhibitory activity from the selected ligands, amino acids involved in its van der Waals interaction with receptor molecules are His249, His250, His253, Ser154, Thr238, Arg245, and Lys246, while those in hydrophobic interactions include Ile241 and Ala242 and one pi-donor hydrogen bond with His249 (Fig. 2).

Docking screening of compounds with Molegro Virtual Docker (MVD)

The standards docked drugs were listed depending on their MolDock score as given in Table 4, and the nature of interaction with the protein structure is shown in Fig. 3. The best pose of the docked molecules was analyzed based on the lowest MolDock score. Docking results showed that ceftriaxone was best docked with the

Table 3 The results of the 58 salicylidene acylhydrazides derivatives with iGemDock v2.1

No	Ligand	Total energy	VDW	HBond	Elec	AverConPair
1	6e7e-44-0.pdb	-87.3827	-87.3827	0	0	18.4138
2	6e7e-43-0.pdb	-78.9421	-78.9421	0	0	19.04
3	6e7e-4-2.pdb	-78.3402	-78.3402	0	0	20.3478
4	6e7e-48-1.pdb	-77.9572	-77.9572	0	0	19.2308
5	6e7e-38-1.pdb	-76.561	-76.561	0	0	17.25
6	6e7e-54-0.pdb	-75.6408	-75.6408	0	0	18.92
7	6e7e-40-1.pdb	-74.12	-74.12	0	0	19.2609
8	6e7e-29-0.pdb	-73.4891	-73.4891	0	0	20
9	6e7e-45-1.pdb	-73.338	-73.338	0	0	18.8889
10	6e7e-45-1.pdb	-73.338	-73.338	0	0	18.8889
11	6e7e-5-2.pdb	-72.2371	-72.2371	0	0	19.0455
12	6e7e-14-1.pdb	-71.4777	-71.4777	0	0	20.3636
13	6e7e-6-1.pdb	-71.3965	-71.3965	0	0	19.7143
14	6e7e-39-0.pdb	-71.2635	-71.2635	0	0	17.0769
15	6e7e-46-1.pdb	-71.1864	-71.1864	0	0	16.4231
16	6e7e-57-1.pdb	-70.8057	-70.8057	0	0	20.1905
17	6e7e-13-0.pdb	-70.6842	-70.6842	0	0	19.4348
18	6e7e-1-2.pdb	-70.1351	-70.1351	0	0	18.2174
19	6e7e-34-0.pdb	-70.0374	-70.0374	0	0	18
20	6e7e-30-2.pdb	-69.8395	-69.8395	0	0	20.4091
21	6e7e-12-0.pdb	-69.6612	-69.6612	0	0	18.7083
22	6e7e-47-0.pdb	-69.4781	-69.4781	0	0	16.88
23	6e7e-28-1.pdb	-69.0599	-69.0599	0	0	17.4167
24	6e7e-41-1.pdb	-68.9204	-68.9204	0	0	20.05
25	6e7e-22-1.pdb	-68.7563	-68.7563	0	0	19.5238
26	6e7e-16-0.pdb	-67.8887	-67.8887	0	0	18.5652
27	6e7e-17-1.pdb	-67.8206	-67.8206	0	0	16.3478
28	6e7e-33-0.pdb	-67.7813	-67.7813	0	0	17.1739
29	6e7e-23-1.pdb	-66.7408	-66.7408	0	0	20.4737
30	6e7e-57-0.pdb	-66.5272	-66.5272	0	0	17.8095
31	6e7e-10-0.pdb	-64.5692	-64.5692	0	0	20.15
32	6e7e-26-2.pdb	-63.4082	-63.4082	0	0	18.2609
33	6e7e-35-0.pdb	-61.2651	-61.2651	0	0	21.6471
34	6e7e-52-2.pdb	-61.1252	-61.1252	0	0	19.3333
35	6e7e-19-0.pdb	-57.6773	-57.6773	0	0	21.4706
36	6e7e-8-0.pdb	-57.1784	-57.1784	0	0	21.3333
37	6e7e-8-0.pdb	-57.1784	-57.1784	0	0	21.3333
38	6e7e-24-1.pdb	-56.8114	-56.8114	0	0	23.5625
39	6e7e-37-0.pdb	-38.9493	-38.9493	0	0	13.2258
40	6e7e-58-0.pdb	-31.2029	-31.2029	0	0	13.3182
41	6e7e-21-0.pdb	-9.41303	-9.41303	0	0	12.375
42	6e7e-15-0.pdb	-1.74972	-1.74972	0	0	10
43	6e7e-50-0.pdb	0.890899	0.890899	0	0	14.0417
44	6e7e-18-0.pdb	11.5326	11.5326	0	0	17.625
45	6e7e-27-0.pdb	18.0226	18.0226	0	0	18.2222
46	6e7e-31-0.pdb	29.5633	29.5633	0	0	12.8846
47	6e7e-2-0.pdb	30.294	30.294	0	0	17.2
48	6e7e-9-0.pdb	56.4153	56.4153	0	0	16.5455
49	6e7e-9-0.pdb	56.4153	56.4153	0	0	16.5455

Table 3 (continued)

No	Ligand	Total energy	VDW	HBond	Elec	AverConPair
50	6e7e-51-0.pdb	70.1743	70.1743	0	0	16.1481
51	6e7e-42-0.pdb	91.6956	91.6956	0	0	13.8889
52	6e7e-25-0.pdb	98.3126	98.3126	0	0	15.44
53	6e7e-11-0.pdb	143.872	143.872	0	0	22.2917
54	6e7e-53-0.pdb	144.199	144.199	0	0	21.4583
55	6e7e-36-0.pdb	181.578	181.578	0	0	16.2
56	6e7e-20-0.pdb	195.275	195.275	0	0	36.8696

protein structure. Ceftriaxone with the protein was found to be the most active with the best MolDock value as (-124.659) as compared to all the other poses as shown in Table 4. The result showed that ceftriaxone perfectly binds with the active regions of the target protein by forming hydrophobic interactions with Gln114, Lys115, and Phe108 as shown in Fig. 3. Oxytetracycline makes other side interactions (unfavorable bump and unfavorable donor–donor) with Gly144, Phe145, and Ser111 amino residue, as such can lead to side effects.

Accordingly, all the 58 salicylidene acylhydrazides derivatives were docked into the binding pocket of the protein. The MolDock score, rerank score, and hydrogen bond values of the five most active compounds from the 58 datasets are provided in Table 5. It was observed (Table 4) that ceftriaxone, in particular, showed a superior binding affinity with a MolDock score of -124.659 , in comparison with compounds 16, 23, 32, 44, and 51 with a MolDock score of -102.325 , -105.42 , -103.832 , -101.187 , and -104.935 , respectively. Hydrogen bond, electrostatic, and hydrophobic interactions of compounds 16, 23, 32, and 51 with the protein are illustrated in Fig. 4. Likewise, docking pose analysis of four best-performing derivatives (compounds 16, 23, 32, and 51) showed the similarity of binding mode compared to the standard drugs. The similarity is observed in the urea moiety of the compounds, which is with Lys160, Asn161, Ser162, Glu158, Met166, Glu248, His252, His251, His249, Thr163, Arg245, and Asp157, respectively. The MolDock score of compounds 16, 23, 32, 44, and 51 is far better than the rest of the standard drugs (ampicillin, oxytetracycline, and chlortetracycline).

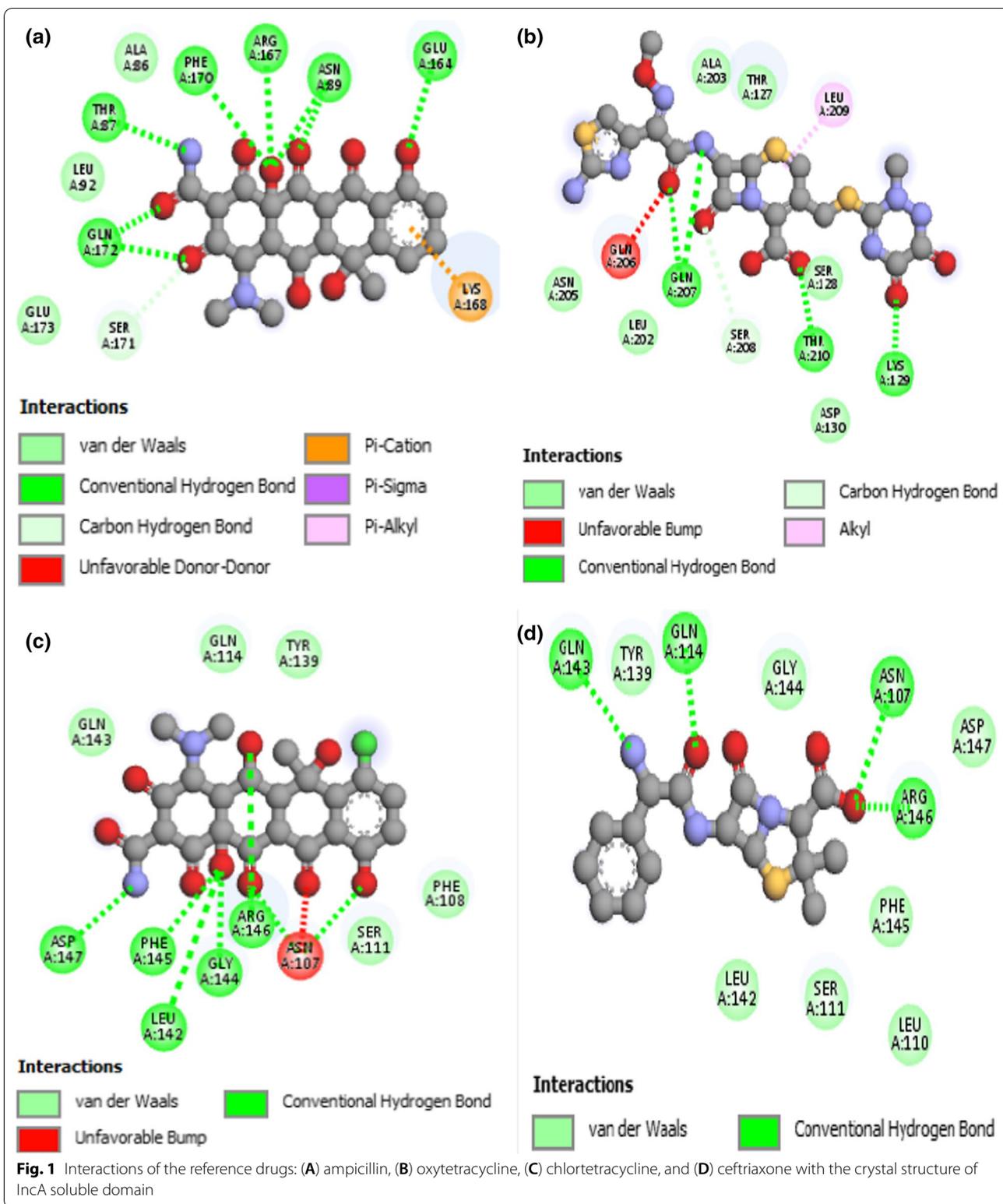
Docking screening of compounds with AutoDock Vina (PyRx software)

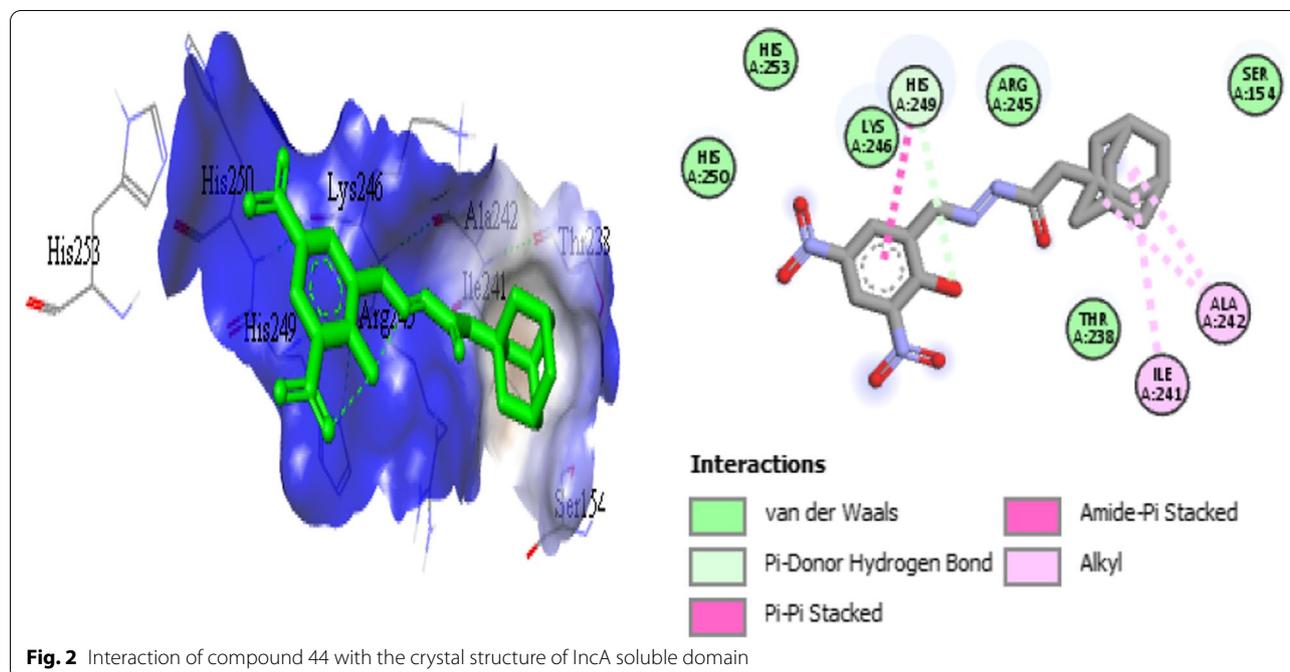
Based on AutoDock Vina (Table 5), out of 58 compounds used in the present study, compounds 36, 37, and 39 exhibited the lowest binding affinity with the enzyme (-6.9 kcal/mol) active sites, followed by compounds 44 and 50 (-6.8 kcal/mol) and compounds 11 and 20

(-6.7 kcal/mol), respectively. Compounds 36, 37, and 39 display the highest docking scores when compared with the standard drugs (Table 4); furthermore, chlortetracycline (standard drug) showed the highest binding energy of -6.8 kcal/mol and exhibited two binding interactions; van der Waals interaction with His249, His252, Thr163, Glu248, and Tyr159 and four hydrogen-bonding interactions with Glu158, Arg245, Asp157, and Asn161 (Fig. 5). Ampicillin, oxytetracycline, and ceftriaxone showed binding affinities of -6.0 , -6.5 , and -6.1 kcal/mol, respectively. These compounds were bound at the van der Waals, conventional and carbon–hydrogen bond, ampicillin has one hydrophobic interaction with Lys235, and ceftriaxone has unfavorable donor–donor with Arg146 as shown in Fig. 5.

The selected compounds (36, 37, 39, and 44) all have a higher binding affinity than the standard drugs except compounds 44 and 50 that have their binding energy equal to chlortetracycline of the standard drug. Compounds 36 and 39 were found to have the highest binding affinity as such are more compatible with the receptor than compounds 37 and 44 with unfavorable bumps (Fig. 6). As illustrated in Fig. 6, there are a variety of interactions between these chemicals, and the target protein revealed that compound 36 interacts with one hydrogen bond with Glu215 and one carbon–hydrogen bond with Phe191. In addition to the binding interactions of compound 36 are hydrophobic (Ile219, Phe191, and Pro194), van der Waals (Thr218, Leu195, and Glu198), and Halogen (Glu188) interaction between amino acid residues, respectively. Compound 39 has three conventional hydrogen bonds with His204, Arg200, and Glu197, one carbon–hydrogen bond with Phe124, and hydrophobic interactions with Arg201, His204, His204, Arg200, and Phe124, respectively.

It was concluded from the docking simulation that compound 36 and compound 39 were strongly bonded through van der Waals, hydrophobic, and conventional



**Table 4** The results of the standard drugs with MVD software

SD	MolDock score	Rerank score	HBond	Ligand	Binding affinity
Ampicillin	-87.3309	-68.6287	-13.8777	Ampicillin	-6.0
Oxytetracycline	-50.6776	-11.9821	-6.89977	Oxytetracycline	-6.5
Chlortetracycline	-50.1173	-55.3497	-9.28234	Chlortetracycline	-6.8
Ceftriaxone	-124.659	-90.9047	-8.66981	Ceftriaxone	-6.1

SD, standard drugs

hydrogen bond interactions and stabilized into the active site of the target protein.

Oral bioavailability analysis of the selected compounds and standards

These findings prompted us to evaluate the physico-chemical properties of our selected lead compounds from the docking tools, one from iGemDock (compound 44), five from MVD (compounds 16, 23, 32, 44, and 51), and five from AutoDock Vina with PyRx (compounds 36, 37, 39, 44, and 50), respectively (Table 6), to identify compounds with optimal oral absorption properties and to guide future structural modifications to improve ADMET properties. The oral bioavailability analysis of selected compounds was predicted using DataWarrior v5.5.0 and is presented in Table 6; following Lipinski's rule of five [25] and Veber's rules of two [26], any compound violating more than one of these rules could have problems related to its bioavailability.

From the table (Table 6), all the compounds can easily pass through the cell membrane since their molecular weight is less than 500 Da, except chlortetracycline that has a molecular weight of more than 500 Da. If the size increases, it will create barriers such as the prevention of passive diffusion through the tight aliphatic side chains of the bilayer membrane [27]. According to the Lipinski rule, cLogP should be less than or equal to 5, all the selected compounds have positive cLogP (affinity for lipids), so they have a good permeability through a biological membrane. Compound 36 is more than the threshold value of 5, while the standard drugs have a negative value of cLogP, which means they are hydrophilic (strong affinity for water) [28]. A drug molecule is expected to be in an aqueous solubility, range of -1 to -5 [29], and the cLogS values of all the selected compounds and standards fall within the range, indicating that the compounds have good absorption and distribution potential except compound 36 with cLogS value of -6.523. The number of hydrogen bond donors

Table 5 The MolDock scores and AutoDock Vina binding affinity of the docked compounds

Positions	Ligand	MolDock score	Rerank score	HBond	AutoDock vina: ligand	Binding affinity
1	6e7e-23	-105.42	-83.3015	-2.3003	6e7e_36	-6.9
2	6e7e-51	-104.935	-86.2557	-0.4690	6e7e_37	-6.9
3	6e7e-32	-103.832	-81.4928	-5.9129	6e7e_39	-6.9
4	6e7e-16	-102.325	-81.2418	-9.2016	6e7e_44	-6.8
5	6e7e-44	-101.187	-81.2352	-2.8332	6e7e_50	-6.8
6	6e7e-0	-101.058	-78.2991	-1.1185	6e7e_11	-6.7
7	6e7e-35	-100.81	-74.1065	-2.7562	6e7e_20	-6.7
8	6e7e-48	-100.468	-83.5565	-4.8169	6e7e_38	-6.6
9	6e7e-27	-99.6987	-47.8776	-5.9033	6e7e_4	-6.5
10	6e7e-15	-99.0472	-59.3321	-7.1583	6e7e_13	-6.5
11	6e7e-41	-97.9657	-83.8199	-8.4460	6e7e_17	-6.5
12	6e7e-34	-97.7763	-75.7789	-2.5000	6e7e_29	-6.5
13	6e7e-31	-97.7655	-71.8349	-4.6374	6e7e_30	-6.5
14	6e7e-29	-97.5903	-80.7864	-1.3863	6e7e_32	-6.5
15	6e7e-25	-97.1211	-76.7611	-4.3095	6e7e_33	-6.5
16	6e7e-5	-97.0019	-78.9432	-2.5118	6e7e_43	-6.5
17	6e7e-43	-96.9476	-65.1119	-2.1904	6e7e_6	-6.4
18	6e7e-55	-96.5008	-80.0093	-4.9261	6e7e_16	-6.4
19	6e7e-42	-96.1401	-61.68	-2.5000	6e7e_21	-6.4
20	6e7e-30	-96.0124	-79.8537	-1.9294	6e7e_26	-6.4
21	6e7e-37	-96.0051	-79.1085	-8.6238	6e7e_42	-6.4
22	6e7e-50	-95.9272	-80.5706	-5.0000	6e7e_51	-6.4
23	6e7e-39	-95.494	-73.5631	-10.064	6e7e_1	-6.3
24	6e7e-47	-94.6542	-44.1998	-5.6681	6e7e_18	-6.3
25	6e7e-46	-93.8322	-62.9706	-8.4617	6e7e_2	-6.2
26	6e7e-26	-93.7623	-80.0366	-1.4509	6e7e_5	-6.2
27	6e7e-11	-93.429	-59.9426	-5.2496	6e7e_10	-6.2
28	6e7e-3	-92.5362	-77.3104	-1.9807	6e7e_23	-6.2
29	6e7e-36	-92.5281	-50.0261	-1.8100	6e7e_25	-6.2
30	6e7e-7	-92.0915	-73.6281	-1.6712	6e7e_49	-6.2
31	6e7e-14	-91.8754	-59.7878	-5.9914	6e7e_3	-6.1
32	6e7e-24	-91.383	-75.9929	-1.7595	6e7e_7	-6.1
33	6e7e-13	-90.7777	-78.6832	-1.1915	6e7e_22	-6.1
34	6e7e-1	-89.6732	-73.1549	-2.8683	6e7e_27	-6.1
35	6e7e-21	-88.9992	-71.4621	-3.0701	6e7e_34	-6.1
36	6e7e-10	-88.9228	-76.5735	-3.2483	6e7e_56	-6.1
37	6e7e-28	-88.1999	-54.2836	-2.0728	6e7e_14	-6
38	6e7e-53	-85.632	-70.8852	-3.0808	6e7e_40	-6
39	6e7e-54	-85.2067	-70.4645	-2.6669	6e7e_41	-6
40	6e7e-12	-84.9434	-68.4636	-2.2346	6e7e_48	-6
41	6e7e-8	-84.2814	-72.91	-5.1099	6e7e_54	-6
42	6e7e-6	-84.2788	-76.5631	-5.6531	6e7e_55	-6
43	6e7e-9	-83.9689	-49.0408	-3.0917	6e7e_57	-6
44	6e7e-18	-83.7369	-67.9812	-1.5962	6e7e_19	-5.9
45	6e7e-22	-83.2384	-54.5351	-8.8782	6e7e_28	-5.9
46	6e7e-20	-83.1924	-73.3381	-2.5000	6e7e_46	-5.9
47	6e7e-40	-83.1795	-68.9391	-10.526	6e7e_53	-5.9
48	6e7e-45	-83.109	-59.1662	-7.3442	6e7e_58	-5.9
49	6e7e-19	-82.4367	-64.012	-2.7144	6e7e_9	-5.8

Table 5 (continued)

Positions	Ligand	MolDock score	Rerank score	HBond	AutoDock vina: ligand	Binding affinity
50	6e7e-4	-80.9586	-69.733	-0.8362	6e7e_12	-5.8
51	6e7e-38	-80.7574	-64.5247	-9.0498	6e7e_15	-5.8
52	6e7e-33	-80.4312	-69.4863	-6.9611	6e7e_31	-5.8
53	6e7e-52	-79.6886	-67.2744	-3.2095	6e7e_47	-5.8
54	6e7e-17	-79.4327	-67.8454	-7.1736	6e7e_45	-5.7
55	6e7e-49	-79.0299	-71.0073	-3.8401	6e7e_8	-5.4
56	6e7e-2	-78.0893	-70.0143	-5.0968	6e7e_35	-5.4
57	-	-	-	-	6e7e_52	-5.3
58	-	-	-	-	6e7e_24	-5.1

and acceptors of the selected compounds conforms to the rule of 5 (such as $H-A \leq 10$; $H-D \leq 5$); this shows that the compounds can influence absorption and permeation and also tend to have an increased chance of reaching the market except for the standards (Std2, Std3, and Std4) as shown in Table 6. The two descriptors (1) rotatable bonds ≤ 10 and (2) total polar surface area $\leq 140 \text{ \AA}^2$ are identified as Veber's rule [26] concerning the oral bioavailability of the drug. Compound 44 and the standards (Std2, Std3, and Std4) violate the total polar surface area as shown in Table 6 with their values more than the standard range.

Drug-likeness and lipophilicity analysis of the selected compounds and standards

From Table 7, all the selected compounds having their ligand efficiency (LE) values greater than 0.3 (LE recommended values ≥ 3) are qualified as a hit, while compound 23 is the most potent of all the selected compounds. For other bioactivity parameters such as ligand lipophilicity efficiency (LLE) and ligand-efficiency-dependent lipophilicity (LELP), their recommended values for a hit are ≥ 5 and -10 to 10 , respectively [30–32]. The LLE values were observed for all the selected compounds, and compounds 16, 44, and 51 are within the recommended range. All the selected compounds obey (LELP) recommended value except compound 36 and compound 37 with LELP values of 20.177 and 14.208, respectively (Table 7).

Toxicity analysis of the selected compounds and standards

The toxicity, total surface area, and relative polar surface area analysis of the selected compounds was performed using the DataWarrior package. All the selected compounds have no toxicity risk, except compound 16 which has high mutagenic and low irritant, while compound 32 toxicity risk factors were high in tumorigenic, reproductive effects, and irritant (Table 8).

The overall theoretical ADMET properties justify that compounds 23, 44, and 51 showed potential lead-like features and can be used for further assessment for *Chlamydia trachomatis* treatment.

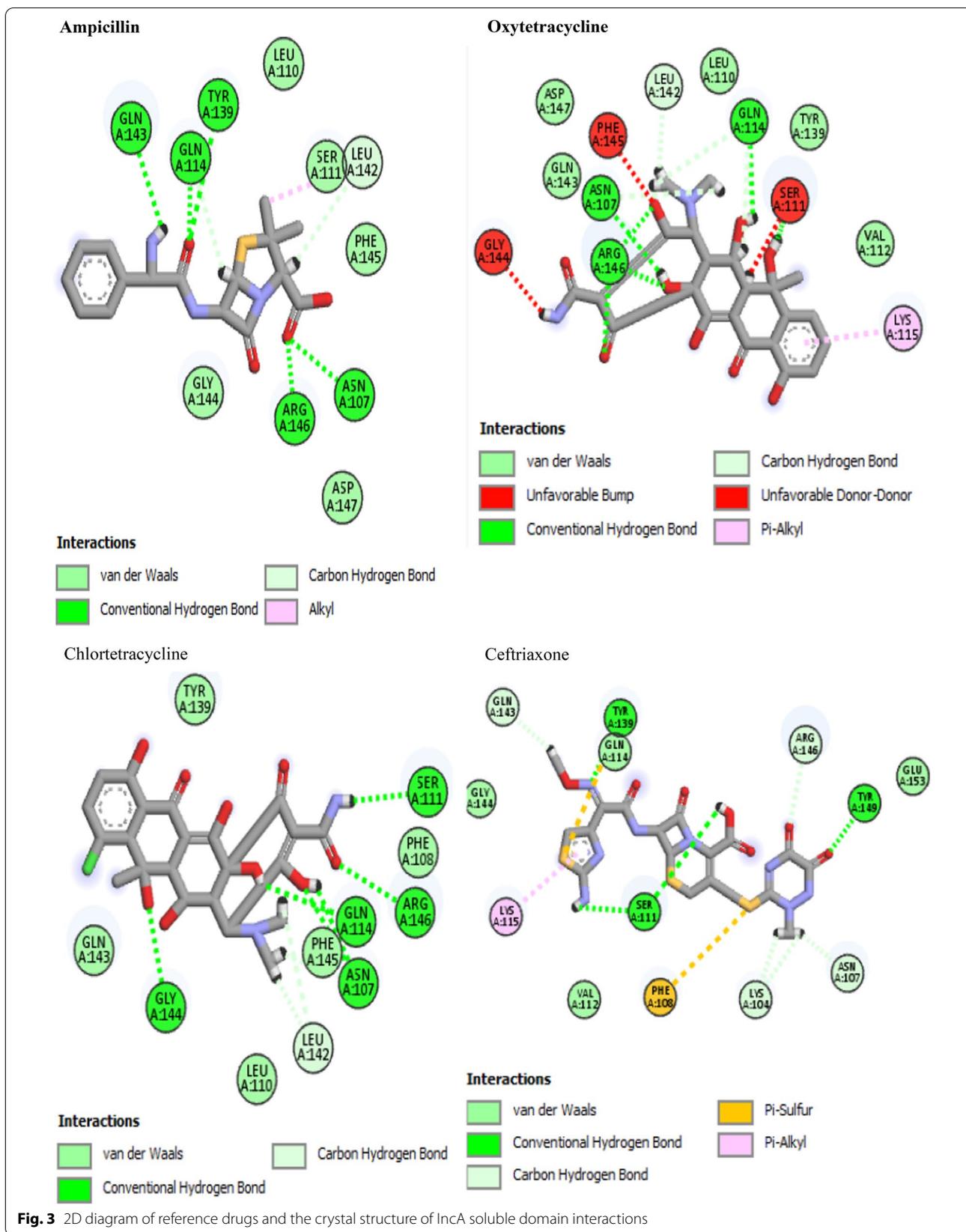
Clearances and computational data designed

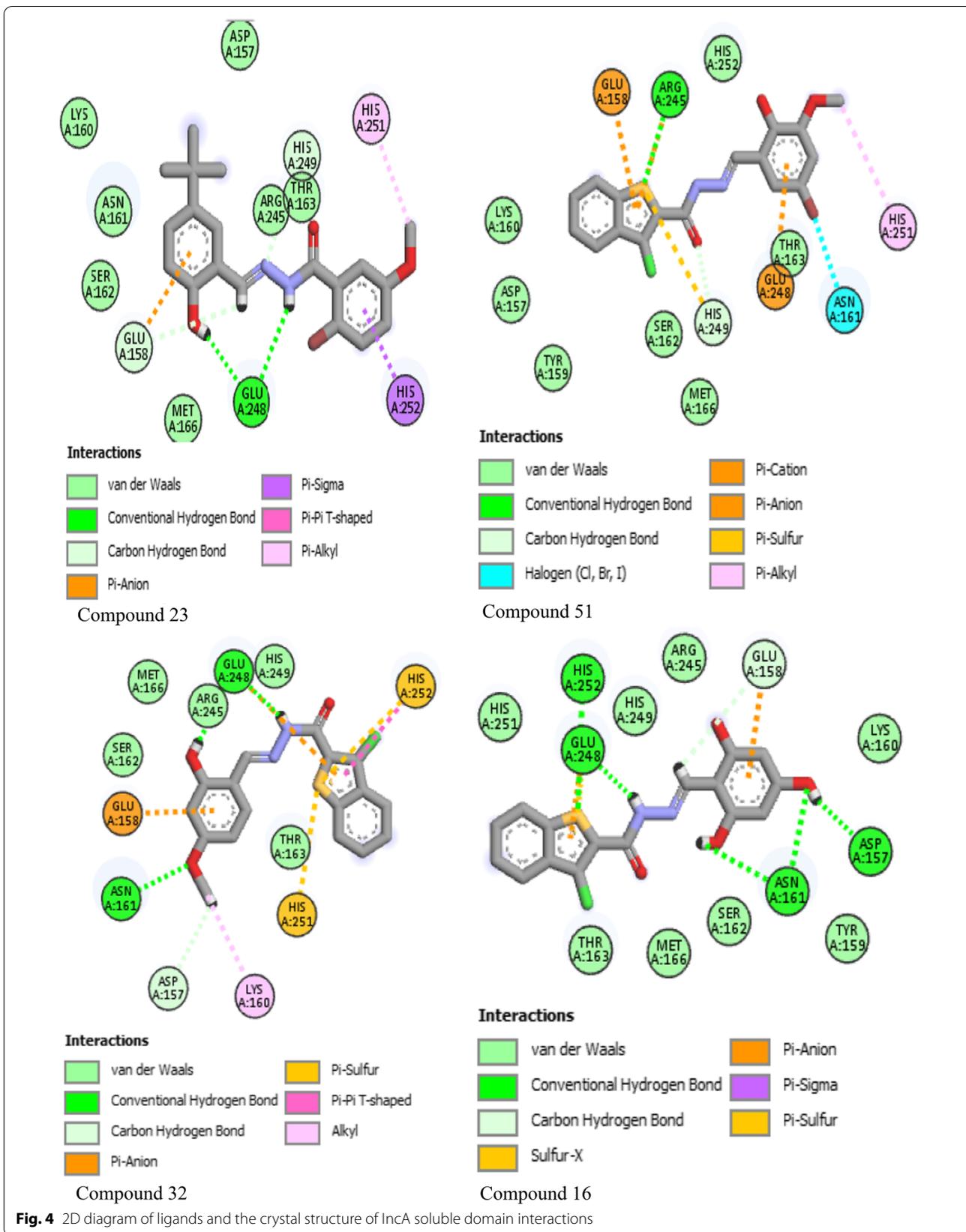
The golden triangle [33] is a visualization tool that helps medicinal chemists find metabolically stable, permeable, and potent drug candidates by combining in vitro permeability, in vitro clearance, and computational data. Moving the design properties into a region with a baseline of $\text{clogP} = -2$ to 5 at $\text{MW} = 200$ and a peak of 450 gives a triangular shape termed the golden triangle, which increases the probability of success in maximizing potency, stability, and permeability. For our selected compounds and standard drugs, it is apparent that compounds 16, 23, 44, and 51 have good permeability and low clearance because they are concentrated within the golden triangle area (Fig. 7). These compounds (16, 23, 44, 51) have the highest probability of success in maximizing potency, stability, and permeability.

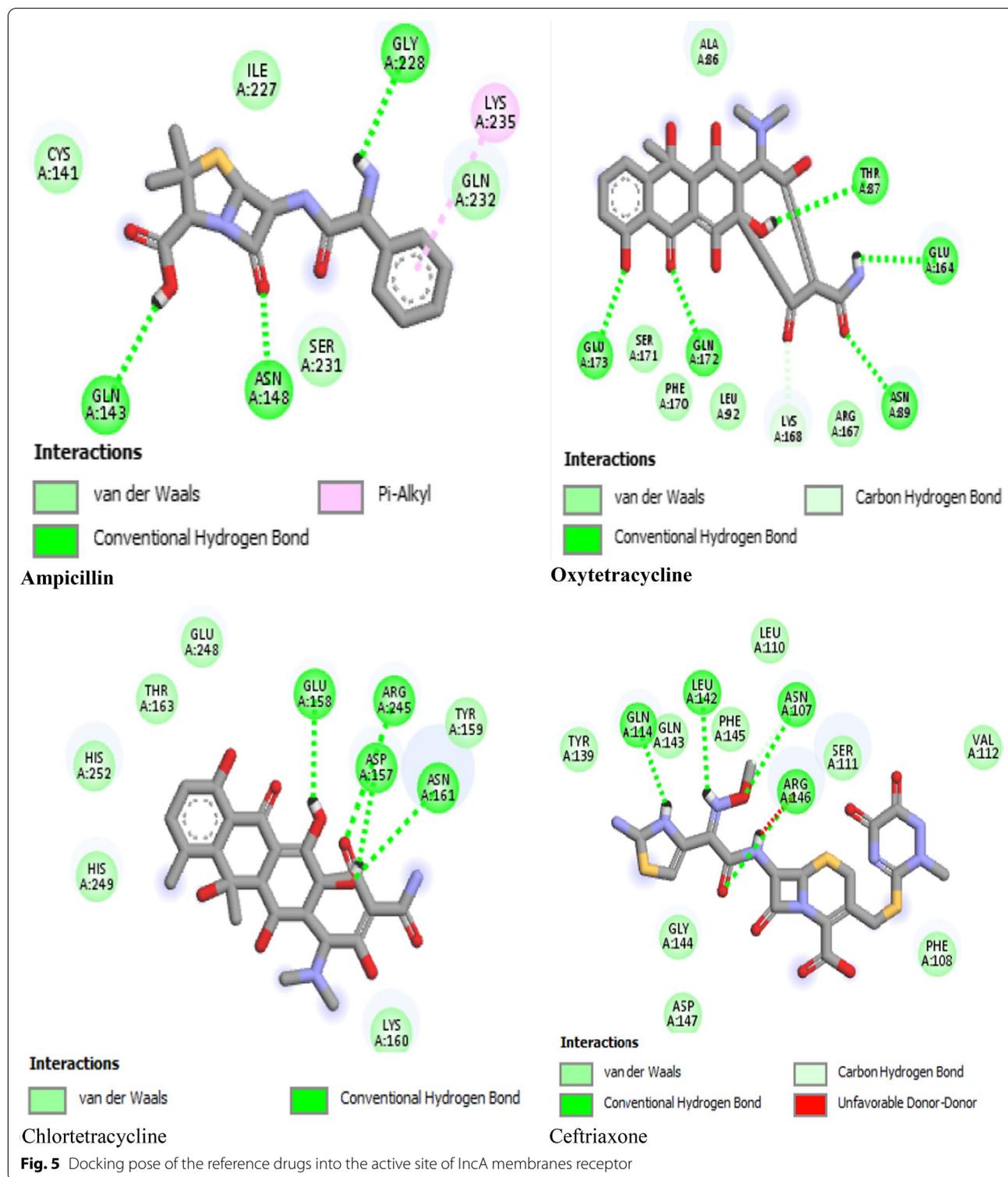
Because they are concentrated within the golden triangle area, compounds 16, 23, 44, and 51 have strong permeability and low clearance for our selected compounds and standard medications (Fig. 7).

Molecular dynamics simulations

The molecular dynamics simulations are widely used to investigate the behavioral and dynamical characteristics of the protein–ligand complex. The MD simulations of the proposed docked ligand were explored for 1 ns. Several parameters such as kinetic energy, total energy, and potential energy were obtained from the MD simulation trajectory. Each of the parameters explains the stability of the complex in dynamics states. Compound 44 came first with iGemdock with a total energy of -87.383 , fifth with MVD with MolDock score of -101.325 , and second with AutoDock Vina with PyRx with a binding

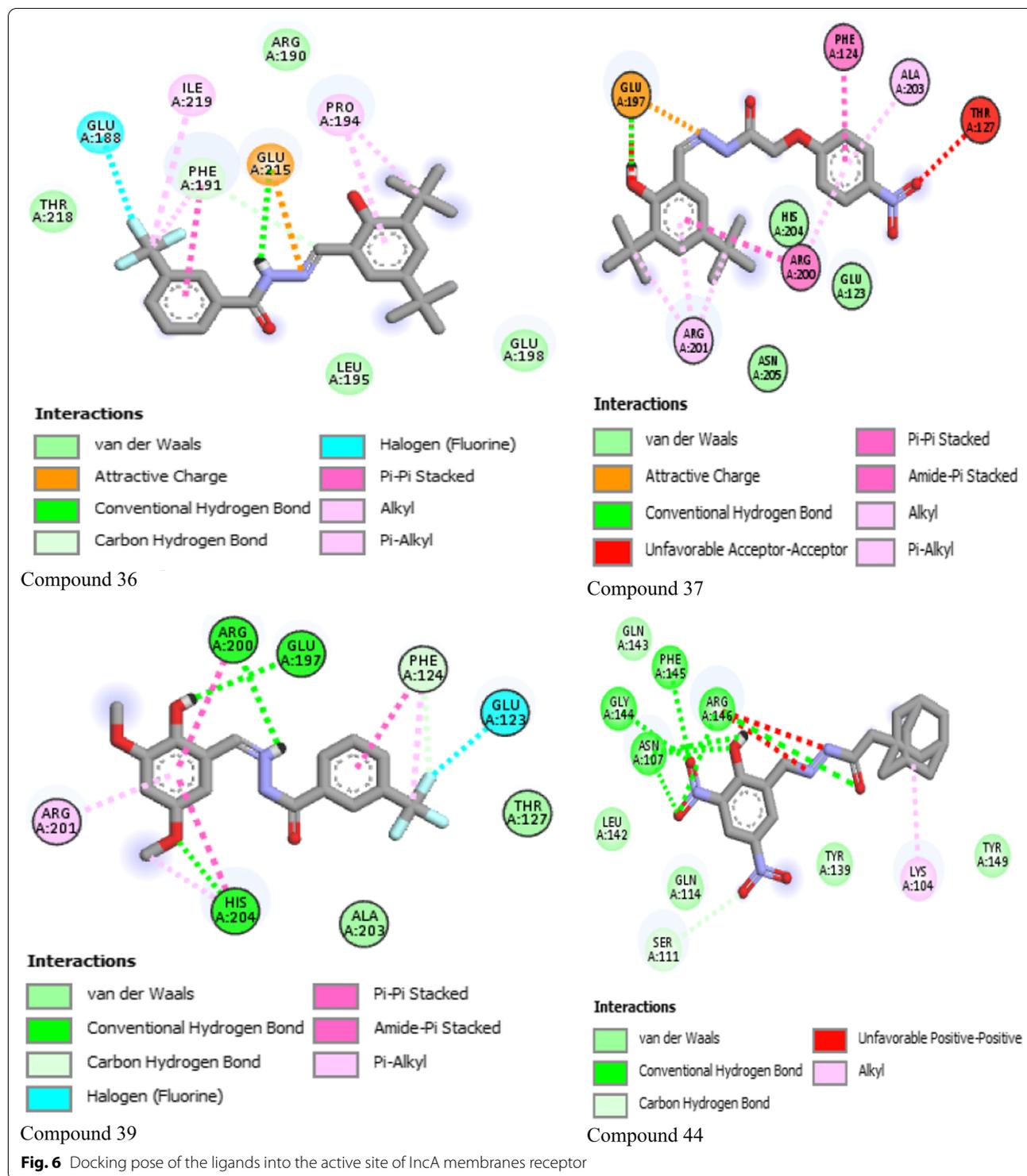






affinity value of -6.8 kcal/mol. These compounds also fit the Lipinski rule of 5 and also Veber rules. Even though compound 44 did not fit with one criterion in Veber's rules (TPSA), it has no mutagenic, tumorigenic, irritant,

and developmental toxicity properties. It also passes lipophilicity indices (LE, LLE, and LELP) [34] and also falls within the golden triangle that was used for 1-ns MD simulations. Figure 8A shows the variation in kinetic



energy versus time in 1 ns of MD simulations. It shows a fluctuation at about 0.89 ns of the MD simulations. In Fig. 8B and C, the total energy and the potential energy versus time in 1-ns MD simulations show a fluctuation

at about 0.8-ns MD simulations. After 500,000 runs (1 ns) using the NAMD package, the kinetic, total, and potential energy values are 51,926.64, -234,493.11, and -286,419.79 kcal/mol, respectively. These proofs show

Table 6 Physicochemical properties of selected compounds in comparison with standard drugs

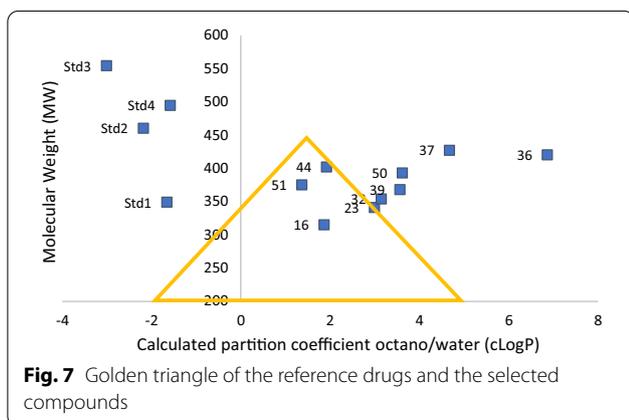
Cpd no	MW	cLogP	cLogS	H-A	H-D	Violation	TPSA	R-Bond
16	315.284	1.864	-3.903	8	2	Nil	116.74	5
23	341.188	2.992	-4.374	6	2	Nil	115.71	3
32	354.148	3.146	-5.357	7	2	Nil	107.51	4
36	420.473	6.868	-6.523	4	2	1	61.69	6
37	427.499	4.673	-5.985	8	2	Nil	116.74	8
39	368.31	3.564	-4.239	6	2	Nil	80.15	6
44	402.406	1.920	-5.6	10	2	Nil	153.33	6
50	393.236	3.618	-4.216	6	3	Nil	91.15	6
51	375.336	1.369	-3.701	10	2	Nil	135.2	8
Std1	349.41	-1.657	-1.565	7	3	Nil	138.02	4
Std2	460.438	-2.184	-1.43	11	7	2	210.85	2
Std3	554.588	-3.011	-2.953	15	4	2	287.82	8
Std4	494.883	-1.578	-2.166	11	7	2	201.85	2

MW = molecular weight; cLogP = calculated logP; sLogS = solubility; H-A = hydrogen bond acceptor; H-D = hydrogen bond donor; TPSA = total polar surface area; R-Bond = rotatable bond; Std1 = ampicillin; Std2 = oxytetracycline; Std3 = chlortetracycline; Std4 = ceftriaxone

Table 7 Lipophilicity analysis of the selected compounds in comparison with standard drugs

Cpd no	DL	LE	LLE	LELP	S-index	MolFle	Mol.Com
16	-0.833	0.465	5.932	4.009	0.565	0.384	0.754
23	-0.855	0.552	4.646	5.425	0.579	0.320	0.682
32	-0.754	0.447	4.349	7.038	0.522	0.387	0.762
36	-6.115	0.340	0.576	20.177	0.467	0.436	0.798
37	-4.090	0.329	2.759	14.208	0.548	0.445	0.759
39	-2.876	0.391	3.845	9.117	0.538	0.444	0.778
44	-0.472	0.348	5.436	5.518	0.483	0.440	0.836
50	1.353	0.417	3.683	8.669	0.667	0.414	0.720
51	-0.898	0.371	5.923	3.695	0.630	0.415	0.732
Std1	9.365	Nil	Nil	Nil	0.542	0.381	0.895
Std2	5.166	Nil	Nil	Nil	0.364	0.328	1.088
Std3	16.694	Nil	Nil	Nil	0.528	0.388	0.959
Std4	5.216	Nil	Nil	Nil	0.353	0.326	1.106

DL, drug-likeness; LE, ligand efficiency; LLE, ligand lipophilic efficiency; LELP, ligand efficiency dependent lipophilicity; S-index, shape index; MolFle, molecular flexibility; Mol.com, molecular complexity; Std1, ampicillin; Std2, oxytetracycline; Std3, chlortetracycline; Std4, ceftriaxone



that energy conservation was satisfied in MD simulations. The binding site residues of compound 44 predicted by MD simulations yield 11 amino acid residues in the binding pocket, namely Ser111, Gln114, Asn107, Leu142, Gly144, Gln143, Lys104, Tyr149, Phe108, Phe145, and Arg146 (Fig. 8D). After a close look at compound 44 complex during MD simulations with the initial molecular docking analysis (Fig. 6), the simulated compound 44 complex adopts a similar binding pocket with the docking complex. The conventional hydrogen bonds, the van der Waals interactions, and the carbon-hydrogen bond observed in the docking complex were monitored. It was observed that the binding interactions of the

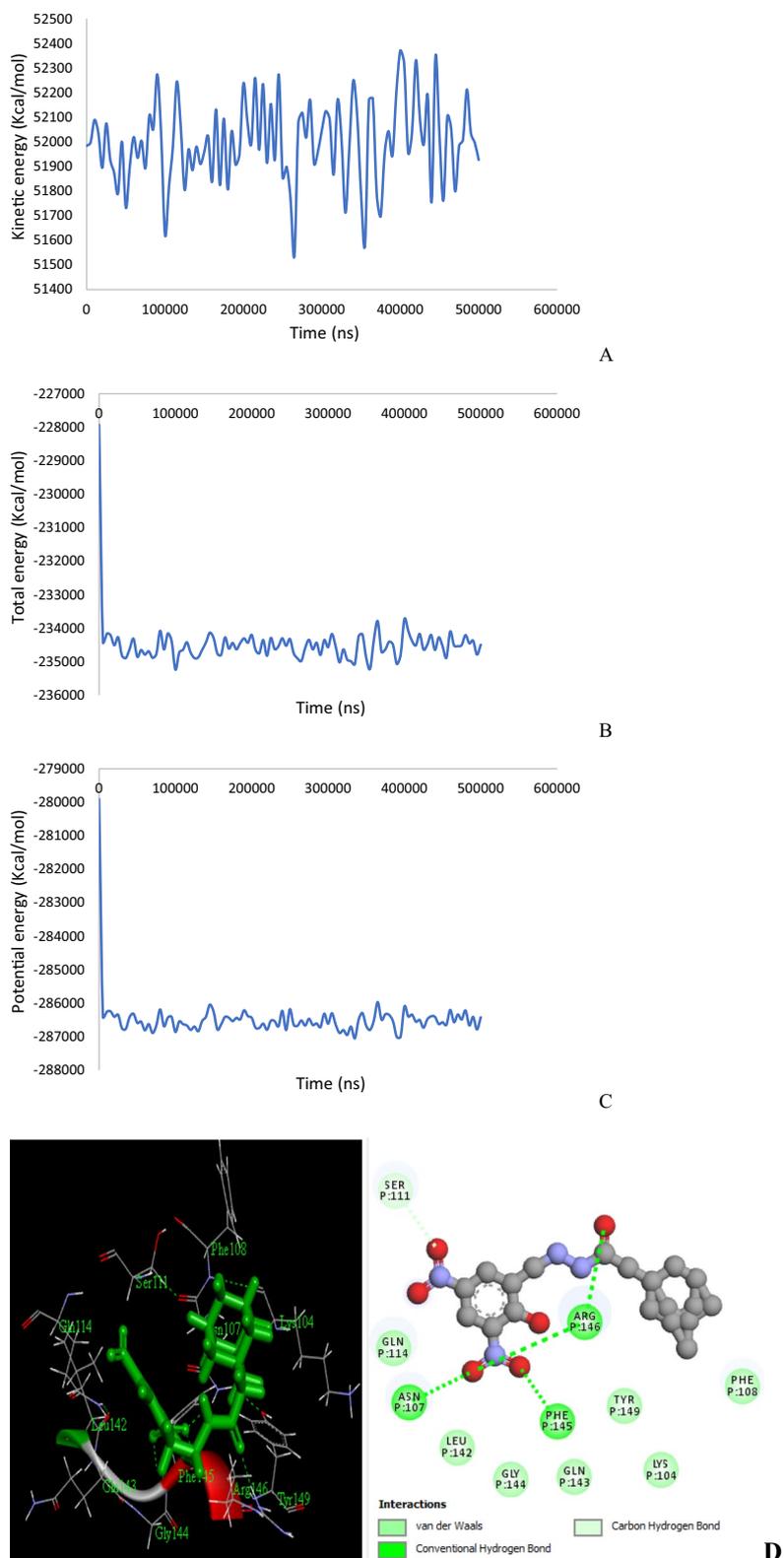


Fig. 8 Interaction of the simulated complex

Table 8 Toxicity risk of selected compounds in comparison with standard drugs

Cpd no	Mutagenic	Tumorigenic	Rep-eff	Irritant	TSA	RPSA
16	High	None	None	Low	239.27	0.374
23	None	None	None	None	217.69	0.420
32	None	High	High	High	247.85	0.321
36	None	None	None	None	317.18	0.155
37	None	None	None	None	335.15	0.267
39	None	None	None	None	267.32	0.259
44	None	None	None	None	284.65	0.386
50	None	None	None	None	268.1	0.269
51	None	None	None	None	285.29	0.384
Std1	None	None	None	None	238.58	0.419
Std2	None	None	None	None	296.37	0.461
Std3	None	None	None	None	365.48	0.610
Std4	None	None	None	None	311.79	0.438

Rep-Eff, reproductive effective; TSA, total surface area; RPSA, relative polar surface area; Std1, ampicillin; Std2, oxytetracycline; Std3, chlortetracycline; Std4, ceftriaxone

docking complex (Fig. 6) were reproduced in MD simulations (Fig. 8D). Compound 44 showed four conventional hydrogen bonds, two with Arg146, one each with Asn107 and Phe145, respectively, and one carbon–hydrogen bond with Ser111. The complex stabilizes its interactions with the binding pocket forming van der Waals interactions with the non-polar surfaces of the active site receptor with the amino acid residues (e.g., Gln114, Gln143, Leu142, Gly144, Lys104, Tyr149, and Phe108).

Conclusions

Herein, molecular docking, ADMET, golden triangle, and MD simulation studies were performed on a series of salicylidene acylhydrazides derivatives against *Chlamydia trachomatis* inhibitors. Three types of powerful docking software (namely MVD, iGemDock, and PyRx) with different algorithms were used to screen the most potent compounds. Docking studies confirm the potential of the nine compounds as novel inhibitors of the *Chlamydia trachomatis* receptor. Besides that, investigated compounds were subjected to the drug-likeness evaluation and golden triangle analysis. The findings revealed that the majority of the compounds would not be orally bioavailable rather would have clearance issues. Compound 44 passes all the quality assurance tests proposed by the model. For further investigation of binding modes of compound 44 and to explain the effects of the compound binding on protein conformation, 1-ns MD simulations were executed. At the end of the MD simulations, a similar position and orientation of the compound in the binding pocket were observed. This significant finding indicated that using MD simulations following ligand docking was beneficial. The nine anticipated salicylidene acylhydrazides inhibitors were predicted through computational techniques; hence,

additional experimental validation would bring fresh insight on these compounds toward designing inhibitory drug molecules against common pathogens of *Chlamydia trachomatis*.

Abbreviations

STD: Sexually transmitted disease; CDC: Centers for Disease Control and Prevention; MD: Molecular dynamics; VMD: Van der Waals; WHO: World Health Organization; ADMET: Absorption, distribution, metabolism, elimination, and toxicity; MW: Molecular weight; cLogP: Calculated LogP; sLogS: Solubility; H–A: Hydrogen bond acceptor; H–D: Hydrogen bond donor; TPSA: Total polar surface area; R-Bond: Rotatable bond; DL: Drug-likeness; LE: Ligand efficiency; LLE: Ligand lipophilic efficiency; LELP: Ligand efficiency-dependent lipophilicity; S-index: Shape index; MolFex: Molecular flexibility; Mol.com: Molecular complexity; Rep-Eff: Reproductive effective; TSA: Total surface area; RPSA: Relative polar surface area; Std1: Ampicillin; Std2: Oxytetracycline; Std3: Chlortetracycline; Std4: Ceftriaxone.

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Authors' contributions

EIE designed and wrote the manuscript, and AU, GAS, and PAM supervised and carried out the statistical analysis. All authors read and approved the final manuscript.

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