

REVIEW

Open Access



Plant bioactive compounds and their mechanistic approaches in the treatment of diabetes: a review

Anshika¹, Rupesh Kumar Pandey^{1*}, Lubhan Singh¹, Sokindra Kumar¹, Prabhat Singh¹, Manish Pathak² and Shruti Jain¹

Abstract

Background: Diabetes mellitus (DM) is a growing disease across the world; diabetes is a complex metabolic disorder in which blood glucose concentration level increases and continue for a prolonged period due to a decrease secretion of insulin or action, resulting in the disorder of carbohydrate, lipid, and protein metabolism. The plant-related bioactive compounds have proven their efficacy with least toxicities and can be utilized for the disease treatment. Our objective is to elucidate the mechanism of action of plant bioactive compounds which can give future direction in diabetes treatment.

Main body: In this review paper, we briefly study more than 200 research papers related to disease and bioactive compounds that have therapeutic applicability in treatment. The plant contains many bio-active compounds which possess in vitro and in vivo anti-diabetic effect which may be responsible for the hypoglycaemic property by inhibiting the digestive enzyme i.e. alpha-amylase and alpha-glucosidase, by producing mimetic action of insulin, by reducing the oxidative stress, by showing antihyperglycemic activity and hypolipidemic activity, by inhibition of aldose reductase, and by increasing or enhancing glucose uptake and insulin secretion.

Conclusion: Our study revealed that terpenes, tannin, flavonoids, saponin, and alkaloids are important bioactive constituents for anti-diabetic activity. The mechanistic approach on alpha-glucosidase and alpha-amylase, hypolipidemic activity, and AR inhibitory action clear-cut explain the therapeutic applicability of these bioactive compounds in disease. Plants that contain these bioactive compounds can be good drug candidates for future research on diabetes treatment.

Keywords: Diabetes mellitus, Medicinal plant, Phytochemicals, Hyperglycaemia, Insulin, STZ, Alloxan

Background

Diabetes is the collection of metabolic illnesses in which increased blood sugar levels persist for a prolonged period due to a malfunction in insulin production that affects the metabolism of various nutrients such as proteins, lipids, and carbohydrates [1]. Metabolism is

normally altered through congenital and environmental variables [2]. The disease pathophysiology suggests that patients may experience frequent urination, thirst, and hunger with other symptoms. Serious complications such as kidney, eye, foot, and another organ failure may be aggravated if properly not managed. In adults, the disease affects 4–5% of people, with the number anticipated to rise to 5.4% by 2025 [3]. Research is now being done on medications that can continually control blood sugar levels. Diabetes is primarily managed with oral hypoglycaemic medications and insulin injections [4]. Injections of hypoglycaemic agents are used in Western medical

*Correspondence: ranu.rupeshpandey@gmail.com

¹ Department of Pharmacology, Kharvel Subharti College of Pharmacy (KSCP), Swami Vivekanand Subharti University, Meerut, Uttar Pradesh, India
Full list of author information is available at the end of the article

treatment for diabetes (insulin, insulin analogs, etc.), the pharmacotherapy suggests that the biguanides, glinides, sulfonylureas and glycosidase inhibitors, thiazolidinedione used as hypoglycaemic medications in oral formulations [5, 6]. These medications have some adverse effects such as severe hypoglycaemia, weight gain, gastrointestinal discomfort, and nausea [7]. Various new medications, including DPP-4 (Dipeptidyl Peptidase) inhibitors, GLP-1 (glucagon-like peptide) analogues, and SGLT-2 (sodium glucose co transporter) inhibitors, have been developed and are available on the market [1]. With the long-term use of oral anti-diabetic agents in patients, the efficacy of all of these inevitably diminishes. As a result, there is an ongoing need to identify and develop novel anti-diabetic medications, particularly given the fact that diabetes has become a global epidemic [2].

Main text

Types of diabetes

The disease has two major types: Type 1 diabetes (T1DM) is an autoimmune disease characterized by insulin insufficiency, whereas Type 2 diabetes (T2D) is characterized by ineffective insulin action [8].

Type 1 diabetes is defined as insulin-dependent diabetes and have characterized by low insulin secretion in the body due to the degeneration of beta cells in the pancreas [9]. Type 1 diabetes patients are always at risk for developing ketoacidosis, the insulin injections required for the maintenance of blood glucose levels under control. The disease is more common in children and teenagers [10].

Causes

1. Genetic factors
2. Environmental factors

Type 2 Diabetes is noninsulin-dependent diabetes due to ineffective insulin action with hyperglycaemia [11–13]. Many of diabetic patients have NIDDM (Non-insulin dependent diabetes mellitus), also known as Type 2 diabetes mellitus (T2DM), which is a lethal disease with severe disability rates.

Causes

1. Obesity
2. Over weight
3. Insulin resistance

Medicinal plants

Medicinal plants and herbs are excellent sources of alternative and complementary medicine and they have a significant function in disease treatment [14]. The

entire medicinal plants specific portions can be used for research purposes in this way.

The plant-derived chemicals are preferred because they were created in a biotic environment and are assumed to have been subjected to evolutionary selection as a result; they communicate better with proteins and are to be prominent medications. Long-term human use of plant extracts provides reliable evidence for diabetes treatment in traditional medicine system. The plant extract contains a variety of phytochemicals that contain many primary and secondary metabolites that can enhance the efficacy of plant-related drugs in treating disease [6].

According to WHO (World Health Organization), to satisfy the primary healthcare needs more than 80% of the world population used natural herbs as a medication furthermore, more than half of all new medications researched and licensed for sale are derived directly from modified medicinal plant products or their active ingredients [15].

Herbal medicine is low in cost, easily available, high effectiveness, and has low side effects due to this feature it is used and prescribed all over the world. As a result, it has been utilized in traditional Indian medicine to treat a variety of ailments and disorders.

Our aim in this review work is to establish the knowledge of plants' bioactive compounds that have the potential to manage diabetes with a full mechanistic approach that can help in future research with regard to efficacy in disease and minimization of toxicities with current allopathic-based medication.

Material and methods

In this review, we systematically reviewed more than two hundred research papers from the Cochrane database. The key words for the search were diabetes, bioactive compounds, plants, and animal models. The exhaustive review was done using the latest information from 2021 and past year data (i.e., 1991), which is relevant to the review work.

Approximately 93 papers were excluded from the review work, of which 57 papers did not have sufficient data related to our work and others did not illustrate significant work.

In the brief review work shown in Table 1, these plants were selected on the basis of their use in the treatment of diabetes in the Indian traditional system of medicine.

Discussion

The various primary and secondary metabolites of the plant are responsible for their activity in diabetes. We conceptualized the available data through a brief literature review. The findings of this work are as follows.

Table 1 Plants potential for anti-diabetic activity

S. no	Plant name	Common name	Part used	Bioactive compound	Animal model	References
1	<i>Berberis cristata</i> L.	Kala Bansha	Seeds	α -Tocopherol, pCumaric acid, Barlerin and Luteolin etc	Alloxan induced diabetic rat	[16]
2	<i>Melia azadirachta</i>	Chinaberry	Leaf	Terpenoids, flavonoids, steriods, acids, anthraquinones, saponins, alkaloids, tannins and limonooids	Alloxan induced diabetic rat	[17, 18]
4	<i>Emblexa officinalis</i> Gaertn.	Indian gooseberry	Leaves	Tannins, amlaic acid, astragalin, ellagic acid, gallo-tannin, kaempferol, kaempferol-3-O-glucoside, phylanthidine, phyllantine and rutin	Streptozotocin-induced type-2 diabetes mellitus (T2DM) rats	[19]
5	<i>Padina boergesenii</i>	Water primose	Whole algae	Pigments, fucoidans, phycocolloids and phlorotannins	STZ induced rat model	[20]
6	<i>Pterocarpus marsupium</i> Roxb.	Indian kino or Bijasar	Bark	(–)-epicatechin, phenolic constituents such as mursupin, protosupin and pierostilbene	Alloxan monohydrate induced male albino wistar rat	[21, 22]
7	<i>Momordica charantia</i> L.	Bitter melons	Fresh or dried fruits	Cucurbitane, charantin, Momordicine II	In STZ-induced T2DM mice	[23]
8	<i>Trigonella foenum-graecum</i> L.	Fenugreek	Fruits	Polyphenols, steriods, lipids, alkaloids, saponins, flavonoids, hydrocarbons, carbohydrates, galactomannanfiber, and amino acids (4-hydroxyisoleucine)	Streptozocin-induced diabetic rats	[24]
9	<i>Hibiscus rosasinensis</i>	China rose	Flower	Tannins, anthraquinones, quinines, phenols, flavonoids, alkaloids, terpenoids, saponins, cardiac glycosides, protein, free amino acids, carbohydrate, reducing sugars, mucilage, essential oils and steriods	Non-obesedabetic (NOD) mouse	[25]
10	<i>Withania somnifera</i> (L.)	Ashwagandha	Root and leaves	Withanolides; withaferin A, withanolide withanolide A, withanolide B, and withanolide IV terpenoids, alkaloids and phenylpropanoids	Streptozotocin induced DM	[26, 27]
11	<i>Ocimum tenuifolium</i>	Tulsi	Fresh or dried leaves	Eugenol, methyleugenol, and – and β -caryophyllene	Alloxaninduced diabetic rats	[28]
12	<i>Tinospora cordifolia</i>	Guduchi	Stem, whole plant	Polyphenol, alkaloid and terpenes	Ehlich ascites tumor cell model system, alloxan treated albino rats	[29, 30]
13	<i>Gymnema sylvestre</i>	Gummar	Leaves	Tannin, quinones,flavonoids, and phenols, saponins (Oleanane and dammarane)	STZ induced male rat model	[31, 32]
14	<i>Ericostemma littorale</i>	Chota-Kirayata	Whole plant	Alkaloids like enicoflavin and gentiocrucine, catechins, saponins, two sterols, triterpenoids, phenolic acid, flavonoids, xanthones and volatile oil	Alloxan-Induced Diabetic Rats	[33, 34]
15	<i>Cinnamomum tamala</i>	Indian Cassia	Whole plant	Saponins, phytosterols, fatty acids, carboxylic acids, monoterpane, sesquiterpene, geraniol, cinnamaldehyde, eugenol	STZ induced male wistar rat model	[34]
16	<i>Allium cepa</i> L.	Onion	Fresh or dried bulb	Quercetin, Rutin, l-Cysteine sulfoxide, Allyl propyl disulphide	Alloxan induced diabetic rats	[35]

Table 1 (continued)

S. no	Plant name	Common name	Part used	Bioactive compound	Animal model	References
17	<i>Azadirachta indica</i> A. Juss	Neem	Dried leaves	Azadirachtins (e.g., azadirachtolide, azadiradione, gedunin and meliacinolin)	STZ induced Mice model	[36]
18	<i>Anacardium occidentale</i> Linn.	Cashew	Leaves	Phenolic, triterpenoids, carbohydrate, xanthoprotein and flavonoids	STZ induced rat model	[37, 38]
19	<i>Brassica oleracea</i>	Broccoli	Sprouts	Antioxidant, vitamins terpenoids, flavonoids, phenolic compound(kaempferol, quercetin), tannins, saponins	Streptozotocin induced diabetic rats	[39]
20	<i>Eugenia jambolana</i>	Indian blackberry or Jamun	Seed	Gallic acid and polyphenolic compounds	Streptozotocin-induced diabetic male albino rat	[40]
21	<i>Cassia auriculata</i> Linn.	Tanner's Cassia	Flowers	Flavonoids, proanthocyanidins and β-sitosterol 8-9	Alloxan Diabetic Rats	[41]
22	<i>Curcuma longa</i>	Turmeric	Rhizomes	Curcumin, diferuloylmethane, curcuminoids, alkaloids, cardiac glycoside and resins, Carbohydrate, terpenes and steroid, Flavonoids, tannins, saponins, balsams and phenols	Alloxan monohydrate induced diabetic rats	[42, 43]
23	<i>Thunbergia laurifolia</i> Linn.	Blue trumpet vine	Flower, leaves	Grandifloric acid, benzyl β-glucopyranoside, benzyl β-(2-O-β-D-glucopyranosyl)-glucopyranoside, 6-C-glucopyranosyl apigenin, 6,8-di-C-glucopyranosyl apigenin, (E)-2-hexenyl-β-glucopyranoside, and hexanol-β-glucopyranoside	Alloxan-induced diabetic rats	[44]
24	<i>Acacia arabica</i>	Babul	Bark	Polyphenols, tannins and flavonoids (for example, quercetin)	streptozotocin-induced diabetic albino rat	[45]
25	<i>Aegle marmelos</i> (L.)	Bael	Leaves	Carotenoids, phenolic, alkaloids, pectins, tannins, coumarins, flavonoids and terpenoids	Streptozotocin-induced diabetic Rats (Long Evan)	[46, 47]
26	<i>Agrimonia eupatoria</i>	Common Agrimony	Leaves	TANNINS, volatile oil and coumarin, gum, phytosterol, polysaccharides,	STZ-diabetic mice	[48]
27	<i>Allium sativum</i> L.	Garlic	Bulb	Flavonoid(quercetin), di (2-propenyl) disulfide and 2-propenyl propyl disulphide	Streptozotocin-induced diabetic wistar rats	[49]
28	<i>Aloe barbadensis</i>	Ghritkumari	Leaves	Aloe emodin, Aloin/Barbaloin	Streptozotocin (STZ)-induced diabetic rats	[50, 51]
29	<i>Berberis hispida</i>	Ash gourd	Fruits	Volatile oils, flavonoids, glycosides, saccharides, proteins, carotenes, vitamins, minerals, β-sitosterin and ursolic acid	Streptozotocin-induced diabetic albino rat	[52, 53]
30	<i>Beta vulgaris</i> L.	Beet root	Root juice	Flavonoids, saponins, sterols and triterpenes	Alloxan monohydrate induced diabetes in albino rats	[54, 55]
31	<i>Caesalpinia bonduc</i> Roxb	Fever nut	Seeds	Furanoditerpenes, phytosterin, β-sitosterol, flavonoids, bondoncellin, aspartic acid, arginine, citrulline, β-carotene	Alloxan monohydrate induced diabetes in wistar rats	[56]

Table 1 (continued)

S. no	Plant name	Common name	Part used	Bioactive compound	Animal model	References
32	<i>Citrullus colocynthis</i>	Bitter apple or a bitter cucumber	Fruits	Flavonoids alkaloids, saponins, tannins and phenols	Streptozotocin (STZ)-induced diabetic rats	[57]
33	<i>Coccinia indica</i>	Ivy gourd	Leaves	Triterpenoids, alkaloids, steriods, carbohydrates, tannins, flavonoids, coumarins, phenols, resins and quinones	Glucocorticoid Induced Insulin Resistance in Wistar Albino rats	[58]
34	<i>Eucalyptus globulus</i>	Dried eucalyptus leaves	Leaves	Eucalypto (cineol), rutin, terpineol, sesquiterpene, alcohols, aliphatic aldehydes, isoamyl alcohol, ethanol, terpenes and tannins	Streptozotocin (STZ)-induced diabetic mice	[59]
35	<i>Ficus bengalensis</i>	Bargad or Banyan tree	Fresh aerial roots	flueocyanidin 3-O-beta-Dgalactosyl cellobioside and a dimethoxy ether of leucopelargonidin-3-O-alpha-L-farnnoside	Streptozotocin (STZ)-induced diabetic Female albino Wistar rat	[60]
34	<i>Hibiscus rosasinensis</i>	China-rose, Gurhal	Fresh flowers	Tannins, anthraquinones, quinines, phenols, flavonoides, alkaloids, triterpenoids, saponins, cardiac glycosides, protein, free amino acids, carbohydrate, reducing sugars, mucilage, essential oils and steroids	Alloxan monohydrate induced diabetes in wistar rats	[61, 62]
35	<i>Ipomoea batatas</i>	Sweet potatoes	Leaves	Polyphenol, vitamins, Anthocyaniins, saponin	Alloxan induced diabetic rats	[63, 64]
36	<i>Jatropha curcas</i>	Purging nut	Leaves	Alkaloids, tannin, flavonoids, saponins, and triterpenoids	Alloxan-induced diabetic rats	[65]
37	<i>Mangifera indica</i> L.	Mango	Leaves	Rhamnetin, catechin, epicatechin, gallic acid derivatives, mangiferin and triflophenone 3-C-β-D-glucoside, flavonoids and polyphenols	Swiss albino mice	[66]
39	<i>Morus alba</i> L.	Mulberry	Branch	Terpenoids, alkaloids, flavonoids (including chalcones and anthocyaniins), phenolic acids, stilbenoids	Streptozotocin (STZ)-induced diabetic mice	[67, 68]
40	<i>Mucuna pruriens</i> L.	Velvet Beans or Cowitch	Seeds	Glutathione, gallic acid, and beta-sitosterol, palmitic stearic, oleic, and linoleic acids., alkaloids(3-methoxy-1,1-dimethyl-6,7-dihydroxy-1,2,3,4-tetrahydroquinoline and 3-methoxy-1,1-dimethyl-7,8-dihydroxy-1,2,3,4-tetrahydroquinoline)	Alloxan-induced diabetic rats	[69, 70]
41	<i>Punica granatum</i> L.	Pomegranate	Fruits	Punicalagin anthocyanin, phenolic acids, non-phenolic acids, tannins, and glutenins (4)	Alloxan induced diabetic rats	[71]
42	<i>Vincarssea</i>	Madagascar periwinkle,	Whole plant	Alkaloids (vinacristine and vinblastine) and tannins	Alloxan induced diabetic rats	[72]
43	<i>Actinidia deliciosa</i>	Kiwi	Fruit	Triterpenoids, saponins, and phenolic compounds (flavonoids, polyphenols, anthraquinones, and coumarins	Alloxan induced diabetic rats	[73]

Table 1 (continued)

S. no	Plant name	Common name	Part used	Bioactive compound	Animal model	References
44	<i>Annona squamosa</i> L.	Custard apple	Leaves	Steroids, alkaloids, saponins, terpenes, tannins, phenolic substances, carbohydrates, volatile oil and mucilage	Streptozotocin (STZ)-induced diabetic Wistar rats	[74]
45	<i>Camellia sinensis</i>	Green tea	Leaves	Alkaloid, flavonoids, tannin	Alloxan induced diabetic rats	[75]
46	<i>Boerhaavia erecta</i> L.	Tar vine	Whole plant	Flavonoids, tannins and glycosides	Streptozotocin-induced diabetic Wistar rats	[76]
47	<i>Agaricus bisporus</i>	Button mushroom	Fruit	Proteins, amino acids, polyphenols, polysaccharides, ergothionins and vitamins	Alloxan-Induced Diabetic Rats	[77]
48	<i>Semecarpus anacardium</i> (L.)	Marking nut tree	Tem bark	Phenolic compounds as carboxylic acid derivatives, bhtlawanols, sterols, glycosides, bhilawanol, anacardic acid, anacardoside and flavonoids	Alloxan induced diabetic Long Evans rats	[78]
49	<i>Spondias mombin</i>	Hog plum	Leaves	Phenolics, sterols, triterpenes, saponins, essential oils, amino acids, and polysaccharides	Alloxan induced diabetic Wistar albino rats	[79, 80]
50	<i>Ruellia tuberosa</i> Linn	Fever root,	Whole plant	Triterpenoids and flavonoids (apigenin, luteolin, 3, 5-diglucoside, apigenin, 7-O-glucuronide,)	Alloxan-Induced Diabetic Rabbits	[81]
51	<i>Sida rhombifolia</i>	Bala, Mahabala	Aerial part	Flavonoids, Alkaloid, flavonoids, tannin, glycosides, Phenols		[82]

Saponin

According to some researchers, the root and bark of *Berberis vulgaris* Linn. show a hypoglycaemic effect due to the presence of saponin which has a stimulating effect on remnant beta cells, along this it improves the lipid profile, so it is used in the diabetes treatment [83]. Fenugreks also contain saponin which inhibits cholesterol absorption and reduce sugar level [84].

Tannin

Many studies indicate that black tea contains many active compounds out of which many are tannins which are 90% catechins that show anti-diabetic action by inhibiting intestinal glucose absorption [85, 86].

Grapes contain epicatechin as a major active compound, which prevents hyperglycaemia by inducing β cell regeneration [87].

Terpenes

Inhibition of α -glucosidase and α -amylase It is reported that a triterpenoid that is heptadienic acid withdrawn from the root of *Potentilla fulgens* inhibits the α -glucosidase enzyme and aids in the treatment of diabetes [88]. A research work reported on stem bark of *Fagara tessmannii* contain Pentacyclic triterpene acetates illustrates inhibition against α -glucosidase [89].

Insulin stimulated action of terpenes The rhizomes of *Costusspeciosus* contain costunolide and help in the management of diabetes by stimulating the restoration of beta cells and producing the insulin resemble action on the peripheral tissue [90].

Action on oxidative stress by terpene compounds Some authors reported that a triterpene known as lupeol found in mango has a significant action in the treatment of diabetes through ROS (Reactive oxygen Species) level and reduced oxidative stress which implicates the antioxidant potential in the liver of Swiss albino mice [91]. A few research works suggests that saffron contain an essential oil known as safranal, a monoterpane that protects diabetic rat against oxidative damage [92]. The reduced blood glucose level and improved antioxidant activity reported by some authors through administration of safranal intraperitoneally in the diabetic rat in a dose-dependent manner [93].

Anti-hyperglycaemic activity of terpene compounds An unsaturated triterpene presents in the root and bark of *Bumelia sartorum* isolated from its ethanolic extraction shows hypoglycaemic action by increased insulin production from β -cells [94]. A clinical trial on *Momordica charantia* reported anti-hyperglycemic activity [95].

Hypolipidemic activity of terpenes A triterpene called momordicoside, which is extracted from the bitter melon *Momordica charantia*, improved the fatty acid oxidation and glucose excretion in both types of mice i.e., insulin sensitive and insulin resistant mice during the OGTT [96]. Some authors reported hypo-lipidemic activity through Momordicoside which stimulates the GLUT4 translocation with increased activity of AMP-activated protein [97].

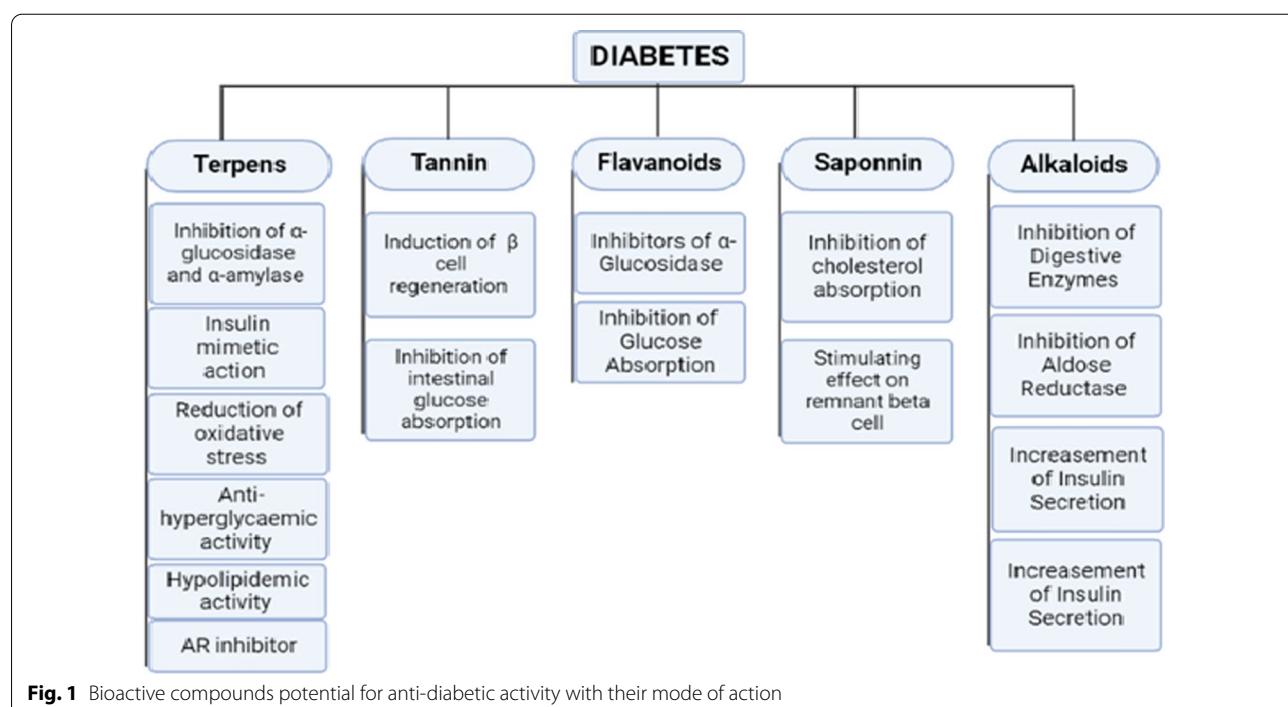
Terpenes as AR inhibitor Research work reported on rat lens implicated the inhibitory effect on AR (Aldose reductase) through friedelane type triterpene salasones A, B, and C and norfriedelane type triterpene, salaquinone A and acylated eudesmane type sesquiterpene, salasol A, which is extracted from *Salacia chinensis* and *Salvia iltiorrhiza* [98, 99].

Alkaloids

Inhibition of digestive enzymes Some researchers revealed that two digestive enzymes hydrolyse the dietary polysaccharides and increase the levels of blood glucose which is α -Amylase present in the pancreatic juice and saliva catalyzes the hydrolysis of α -1,4-glycosidic linkages of starch, glycogen, and various oligosaccharides and increase the blood glucose level and the second one is α -Glucosidase secreted by cells lining in the epithelial cells of the small intestine catalyzes the hydrolytic breakdown of oligosaccharides into absorbable monosaccharides and causes postprandial hyperglycaemia. Alkaloids inhibit these digestive enzymes and decrease the post-prandial blood glucose level [100].

Inhibition of aldose reductase and protein tyrosine phosphatase-1B The compound which has both antioxidant and AR inhibitory activities piqued the interest of the scientific community for research to manage diabetes [101]. In hyperglycaemic conditions, AR increases the sorbitol and its metabolite accumulation in a cell leading to osmotic swelling, overproduction of reactive oxygen species, and cell dysfunction [102].

Effect on insulin secretion Various researchers have concluded that for the rise of insulin release and decrement of blood sugar concentration, inhibition of DPP-IV is required because diminishing DPP-IV enhances glucose tolerance because of latencies in the action of GLP-1 and GIP [103]. GLP-1 and GIP are two hormones that stimulate insulin release [104], decrease glucagon release, improve glucose digestion increased lipoprotein lipase activity and regulate fatty acid production and enhance β -cell proliferation and cell survival [105].



Enhancement of glucose uptake Vindolicine III an alkaloid isolated from the *Catharanthus roseus* (L.) is beneficial in the treatment of hyperglycaemia because it increases glucose absorption through translocation of glucose transporter 4 (GLUT-4) [106].

Flavonoids

Inhibition of α-glucosidase Flavonoid reduces the postprandial blood sugar concentration by inhibiting the α-Glucosidase enzyme, an enzyme present in the small intestine epithelium and involved in carbohydrate digestion, by delaying the conversion of complex carbohydrates to glucose by -glucosidase inhibition, glucose absorption in the small intestine is also delayed, ultimately lowering postprandial blood sugar levels [107].

Inhibiting glucose absorption Type II diabetes GLUT4 plays an important role in homeostasis through glucose uptake mechanism. Diabetes is managed by inhibiting glucose absorption and this can be achieved by inhibiting the GLUT4 translocation [108].

The mechanism of action can be seen in Fig. 1.

Conclusion

Our review work strongly suggests that plant which contains metabolites such as flavonoids, terpenes and alkaloids is having therapeutic value in the treatment of

diabetes. The mechanism of action on alpha-glucosidase and alpha-amylase, hypolipidemic activity, and AR inhibitory action explains that these phyto-constituents can be utilized for future research on diabetes treatment. In our future research work, we will try to emphasize on these bioactive compounds for drug discovery process for diabetes treatment.

Abbreviations

DM: Diabetes mellitus; α-amylase: Alpha amylase; β-amylase: Beta amylase; STZ: Streptozotocin; DPP4: Dipeptidyl peptidase-4; GLP-1: Glucagon-like peptide 1; SGLT-2: Sodium glucose cotransporter-2; T1DM: Type 1 diabetes mellitus; T2DM: Type 2 diabetes mellitus; NIDDM: Non-insulin dependent diabetes mellitus; ROS: Reactive oxygen species; OGTT: Oral glucose tolerance test; GLUT4: Glucose transporter type 4; AMP: Adenosine monophosphate; AR: Aldose reductase; GIP: Gastric inhibitory polypeptide.

Acknowledgements

Not applicable.

Author contributions

In the present review, A analyzed the data related to disease and treatment approaches with Bioactive compounds and was the most important contribution in making the manuscript. RKP and LS performed the systematic evaluation of points related to results. SK elaborated on the conclusion. PS, MP and SJ contributed in terpene and alkaloids moa. All authors read and approved the final manuscript.

Funding

Not applicable.

Availability of data and materials

All data and material are available upon request.

Declarations

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Author details

¹Department of Pharmacology, Kharvel Subharti College of Pharmacy (KSCP), Swami Vivekanand Subharti University, Meerut, Uttar Pradesh, India. ²Department of Pharmaceutical Chemistry, Kharvel Subharti College of Pharmacy, Swami Vivekanand Subharti University, Meerut, Uttar Pradesh, India.

Received: 28 September 2022 Accepted: 28 November 2022

Published online: 09 December 2022

References

- Bai L, Li X, He L, Zheng Y, Lu H, Li J, Zhong L, Tong R, Jiang Z, Shi JL (2019) Antidiabetic potential of flavonoids from traditional Chinese medicine: a review. *Am J Chin Med* 47(5):933–957. <https://doi.org/10.1142/S0192415X1950049>
- Zhu Y, Zhao J, Luo L, Gao Y, Bao H, Li P, Zhang H (2021) Research progress of indole compounds with potential antidiabetic activity. *Eur J Med Chem* 223:113665. <https://doi.org/10.1016/j.ejmech.2021.113665>
- Moller DE, Flier JS (1991) Insulin resistance-mechanisms, syndromes, and implications. *N Engl J Med* 325:938–948
- Wang PC, Zhao S, Yang BY, Wang QH, Kuang HX (2016) Anti-diabetic polysaccharides from natural sources: a review. *Carbohydr Polym* 148:86–97
- Grover JK, Yadav S, Vats V (2021) Medicinal plants of India with anti-diabetic potential. *J Ethnopharmacol* 81(1):81–100. [https://doi.org/10.1016/s0378-8741\(02\)00059-4](https://doi.org/10.1016/s0378-8741(02)00059-4)
- Mishra GP, Sharma R, Jain M, Bandyopadhyay D (2021) Syntheses, biological evaluation of some novel substituted benzoic acid derivatives bearing hydrazone as linker. *Res Chem Intermed* 47:5061–5078. <https://doi.org/10.1007/s11164-021-04555-y>
- Mohammed A, Ibrahim MA, Tajudddeen N, Aliyu AB, Isah MB (2020) Antidiabetic potential of anthraquinones: a review. *Phytother Res* 34(3):486–504. <https://doi.org/10.1002/ptr.6544>
- Chen Q, Zhu L, Tang Y, Zhao Z, Yi T, Chen H (2017) Preparation-related structural diversity and medical potential in the treatment of diabetes mellitus with ginseng pectins. *Ann NY Acad Sci* 1401(1):75–89. <https://doi.org/10.1111/nyas.13424>
- Salsali A, Nathan M (2006) A review of types 1 and 2 diabetes mellitus and their treatment with insulin. *Am J Ther* 13(4):349–361. <https://doi.org/10.1097/00045391-200607000-00012>
- Lukmanji Z (2003) Role of nutrition in the management of diabetes mellitus. *Forum Nutr* 56:170–174
- Galicia-Garcia U, Benito-Vicente A, Jebari S, Larrea-Sebal A, Siddiqi H, Uribe KB, Ostolaza H, Martin C (2020) Pathophysiology of type 2 diabetes mellitus. *Int J Mol Sci* 21(17):6275. <https://doi.org/10.3390/ijms21176275>
- Tripathy D, Chavez AO (2010) Defects in insulin secretion and action in the pathogenesis of type 2 diabetes mellitus. *Curr Diab Rep* 10(3):184–191. <https://doi.org/10.1007/s11892-010-0115-5>
- Salehi B, Ata A, Kumar AVN, Sharopov F, Ramirez-Alarcon K, Ruiz-Ortega A, Abdulmajid Ayatollahi SA, Fokou TVP, Kobarfard F, Zakaria AZ, Iriti M, Taheri Y, Martorell M, Sureda A, Setzer NW, Durazzo A, Lucarini M, Santini A, Capasso R, Ostrand AE, Rahaman AU, Choudhary MI, Chao WC, Rad JS (2019) Antidiabetic potential of medicinal plants and their active components. *Biomolecules* 9(10):551
- Khan MF, Rawat AK, Khatoon S, Hussain MK, Mishra A, Negi DS (2018) In vitro and in vivo antidiabetic effect of extracts of *Melia azedarach*, *Zanthoxylum alatum*, and *Tanacetum nubigenum*. *Integr Med Res* 7(2):176–183. <https://doi.org/10.1016/j.imr.2018.03.004>
- Wang W, Xu J, Fang H, Li Z, Li M (2020) Advances and challenges in medicinal plant breeding. *Plant Sci* 298:110573. <https://doi.org/10.1016/j.plantsci.2020.110573>
- Singh R, Rajasree PH, Sankar C (2012) Screening for anti diabetic activity of the ethanolic extract of *Barleria Cristata* seeds. *Int J Pharm Life Sci* 3(10):2044–2047
- Vijayanand S, Wesely EG (2011) Evaluation of antidiabetic activity of *Melia azadirach* on alloxan induced diabetic rats. *Int J Curr Pharm Res* 3:37–40
- Ahmad, Avanapu SR, Shaik R, Ibrahim M (2012) Phytochemical studies and antioxidant activity of *Melia azedarach linn* leaves by DPPH scavenging Assay. *Int J Pharma Appl* 3(1):271–276
- Nain P, Saini V, Sharma S, Nain J (2012) Antidiabetic and antioxidant potential of *Emblica officinalis* Gaertn. leaves extract in streptozotocin-induced type-2 diabetes mellitus (T2DM) rats. *J Ethnopharmacol* 142(1):65–71. <https://doi.org/10.1016/j.jep.2012.04.014>
- Palanisamy S, Sudha S, Prakash S (2014) Antidiabetic activity of aqueous extract of *Padina boergesenii* in streptozotocin-induced diabetic rats. *Int J Pharm Sci* 6:418–422. <https://doi.org/10.20959/wipr20178-9153>
- Dhanabal SP, Kokate CK, Ramanathan M, Kumar EP, Suresh B (2006) Hypoglycaemic activity of *Pterocarpus marsupium Roxb*. *Phytother Res* 20(1):3–8. <https://doi.org/10.1002/ptr.1819>
- Manickam M, Ramanathan M, Jahromi MA, Chansouria JP, Ray AB (1997) Antihyperglycemic activity of phenolics from *Pterocarpus marsupium*. *J Nat Prod* 60(6):609–610. <https://doi.org/10.1021/np9607013>
- Mahmoud MF, El Ashry FE, El Maraghy NN, Fahmy A (2017) Studies on the antidiabetic activities of *Momordica charantia* fruit juice in streptozotocin-induced diabetic rats. *Pharm Biol* 55(1):758–765. <https://doi.org/10.1080/13880209.2016.1275026>
- Baset M, Ali T, Elshamy H, El SA, Sami D, Tawfik BM (2020) Anti-diabetic effects of fenugreek (*Trigonella foenum-graecum*): a comparison between oral and intraperitoneal administration—an animal study. *Int J Funct Nutr*. <https://doi.org/10.3892/ijfn.2020.2>
- Moqbel F, Naik P, Habeeb N, Subramanyam S (2011) Antidiabetic properties of *Hibiscus rosa sinensis* L. leaf extract fractions on nonobese diabetic (NOD) mouse. *Indian J Exp Biol* 49(1):24–29
- Gorelick J, Rosenberg R, Smotrich A, Hanus L, Bernstein N (2015) Hypoglycemic activity of withanolides and elicited *Withania somnifera*. *Phytochemistry* 116:283–289. <https://doi.org/10.1016/j.phytochem.2015.02.029>
- Jena S (2018) Anti-diabetic effects of *Withania somnifera* root and leaf extracts on streptozotocin induced diabetic rats. *J Cell Tissue Res* 13(1):3597–3601
- Jayant S, Srivastava N (2016) Effect of *Ocimum sanctum* against alloxan induced diabetes and biochemical alteration in rats. *Integr Obes Diabetes*. <https://doi.org/10.16761/iod.1000162>
- Joladarashi D, Chilkunda ND, Salimath PV (2014) Glucose uptake-stimulatory activity of *Tinospora cordifolia* stem extracts in Ehrlich ascites tumor cell model system. *J Food Sci Technol* 51(1):178–182. <https://doi.org/10.1007/s13197-011-0480-3>
- Kinkar B, Patil K (2015) Antidiabetic activity of *Tinospora cordifolia* (fam: menispermaceae) in alloxan treated albino rats. *Appl Sci Res* 1(5):316–319
- Khan F, Sarker M, Ming LC, Mohamed IN, Zhao C, Sheikh BY, Tsong HF, Rashid MA (2019) Comprehensive review on phytochemicals, pharmacological and clinical potentials of *Gymnema sylvestre*. *Front Pharmacol* 10:1223. <https://doi.org/10.3389/fphar.2019.01223>
- El-Shafey A, El-Ezabi M, Selim M, Ouda H, Ibrahim D (2013) Effect of *Gymnema sylvestre* R. Br. leaves extract on certain physiological parameters of diabetic rats. *J King Saud Univ Sci* 25:135–141. <https://doi.org/10.1016/j.jksus.2012.11.001>
- Maroo J, Ghosh A, Mathur R, Vasu VT, Gupta S (2003) Antidiabetic efficacy of *Ericostemma littorale* methanol extract in alloxan-induced diabetic rats. *Pharm Biol* 41(5):388–391. <https://doi.org/10.1076/phbi.41.5.388.15943>
- Bisht S, Sisodia SS (2011) Assessment of antidiabetic potential of *Cinnamomum tamala* leaves extract in streptozotocin induced diabetic rats. *Indian J Pharmacol* 43(5):582–585. <https://doi.org/10.4103/0253-7613.84977>

35. Ozougwu J (2011) Anti-diabetic effects of *Allium cepa* (ONIONS) aqueous extracts on alloxan-induced diabetic *rattus norvegicus*. *Pharmacologyonline* 1:270–281
36. Bhat M, Kothiwale SK, Tirmale AR, Bhargava SY, Joshi BN (2011) Anti-diabetic properties of *Azadirachta indica* and *Bougainvillea spectabilis*: in vivo studies in murine diabetes model. *Evid Based Complement Alternat Med.* <https://doi.org/10.1093/ecam/nep033>
37. Ngozika OF, Nnachetam UV, Ndidi OC (2020) Phytochemical and anti-bacterial activities of *Anacardium occidentale* fruits extracts (cashew) on two drug resistant bacteria. *Int J Health Sci* 5(2):81–87
38. Jaiswal YS, Tatke PA, Gabhe SY, Vaidya AB (2016) Antidiabetic activity of extracts of *Anacardium occidentale* Linn. leaves on n-streptozotocin diabetic rats. *J Tradit Complement Med* 7(4):421–427. <https://doi.org/10.1016/j.jtcme.2016.11.007>
39. Sahai V, Kumar V (2020) Anti-diabetic, hepatoprotective and antioxidant potential of *Brassica oleracea* sprouts. *Biocatal Agric Biotechnol.* <https://doi.org/10.1016/j.babc.2020.101623>
40. Jana K, Bera T, Ghosh D (2015) Antidiabetic effects of *Eugenia jambolana* in the streptozotocin-induced diabetic male albino rat. *Biomark Genom Med* 7(3):116–124. <https://doi.org/10.1016/j.bgwm.2015.08.001>
41. Surana SJ, Gokhale SB, Jadhav RB, Sawant RL, Wadekar JB (2008) Anti-hyperglycemic activity of various fractions of *Cassia auriculata* Linn. in alloxan diabetic. *Indian J Pharm Sci* 70(2):227–229. <https://doi.org/10.4103/0250-474X.41461>
42. Olatunde A, Luka CD, Tijjani H, Obidola SM, Joel E (2014) Anti-diabetic activity of aqueous extract of *Curcuma longa* (Linn.) rhizome in normal and alloxan-induced diabetic rats. *Researcher* 6:58–65
43. Den Hartogh DJ, Gabriel A, Tsiani E (2020) Antidiabetic properties of *Curcumin I*: evidence from in vitro studies. *Nutrients* 12(1):18. <https://doi.org/10.3390/nu12010118>
44. Aritajat S, Wutteeraop S, Saenphet K (2004) Anti-diabetic effect of *Thunbergia laurifolia* Linn. aqueous extract. *Southeast Asian J Trop Med Public Health* 35:53–58
45. Hegazy GA, Alnoury AM, Gad HG (2013) The role of *Acacia Arabica* extract as an antidiabetic, antihyperlipidemic, and antioxidant in streptozotocin-induced diabetic rats. *Saudi Med J* 34(7):727–733
46. Manandhar B, Paudel KR, Sharma B, Karki R (2018) Phytochemical profile and pharmacological activity of *Aegle marmelos* Linn. *J Integr Med* 16(3):153–163. <https://doi.org/10.1016/j.joim.2018.04.007>
47. Ansari P, Afroz N, Jalil S, Azad SB, Mustakim MG, Anwar S, Haque SM, Hossain SM, Tony RR, Hannan JM (2017) Anti-hyperglycemic activity of *Aegle marmelos* (L.) corr. is partly mediated by increased insulin secretion, α-amylase inhibition, and retardation of glucose absorption. *J Pediatr Endocrinol Metab* 30(1):37–47. <https://doi.org/10.1515/jpem-2016-0160>
48. Gray AM, Flatt PR (1998) Actions of the traditional anti-diabetic plant, *Agrimonia eupatoria* (agrimony): effects on hyperglycaemia, cellular glucose metabolism and insulin secretion. *Br J Nutr* 80(1):109–114. <https://doi.org/10.1017/s0007114598001834>
49. Eidi A, Eidi M, Esmaeili E (2006) Antidiabetic effect of garlic (*Allium sativum* L.) in normal and streptozotocin-induced diabetic rats. *Phytomedicine* 13(9–10):624–629. <https://doi.org/10.1016/j.phymed.2005.09.010>
50. Rajasekaran S, Sivagnanam K, Ravi K, Subramanian S (2004) Hypoglycemic effect of Aloe vera gel on streptozotocin-induced diabetes in experimental rats. *J Med Food* 7(1):61–66. <https://doi.org/10.1089/109662004322984725>
51. Sanchez M, Gonzalez-Burgos E, Iglesias I, Gomez-Serranillos MP (2020) Pharmacological update properties of aloe vera and its major active constituents. *Molecules* 25(6):1324. <https://doi.org/10.3390/molecules25061324>
52. Sonowal A, Mahatma A, Kumar MS (2015) Evaluation of antidiabetic potential of methanolic extract of *Benincasa hispida* in streptozotocin induced diabetic rats. *Int J Pharm Sci Res* 6(8):3334–3343. [https://doi.org/10.13040/IJPSR.0975-8232.6\(8\).3334-43](https://doi.org/10.13040/IJPSR.0975-8232.6(8).3334-43)
53. Snaif AE (2017) The pharmacological importance of *Benincasa hispida*: a review. *J Pharm Boil* 5(4):240–253
54. Dubey NK, Kumar S, Shachi K, Dubey U (2020) Anti-diabetic and haematinic effects of beet root juice (*Beta vulgaris* L.) in alloxan induced type-1 diabetic albino rats. *J Diabetes Res Ther.* <https://doi.org/10.16966/2380-5544.150>
55. Mirmiran P, Houshialsadat Z, Gaeini Z, Bahadoran Z, Azizi F (2020) Functional properties of beetroot (*Beta vulgaris*) in management of cardio-metabolic diseases. *Nutr Metab.* <https://doi.org/10.1186/s12986-019-0421-0>
56. Kannur DM, Hukkeri VI, Akki KS (2006) Antidiabetic activity of *Caesalpinia bonduc* seed extracts in rats. *Fitoterapia* 77(7–8):546–549. <https://doi.org/10.1016/j.fitote.2006.06.013>
57. Ghauri AO, Ahmad S, Rehman T (2020) In vitro and in vivo anti-diabetic activity of *Citrullus colocynthis* pulpy flesh with seeds hydro-ethanolic extract. *J Complement Integr Med.* <https://doi.org/10.1515/jcim-2018-0228>
58. Koyagura N, Kumar V, Shanmugam C (2021) Anti-diabetic and hypolipidemic effect of *Coccinia Indica* in glucocorticoid induced insulin resistance. *Biomed Pharmacol J* 14:133–140. <https://doi.org/10.13005/bpj/2107>
59. Gray AM, Flatt PR (1998) Antihyperglycemic actions of *Eucalyptus globulus* (Eucalyptus) are associated with pancreatic and extra-pancreatic effects in mice. *J Nutr* 128(12):2319–2323. <https://doi.org/10.1093/jn/128.12.2319>
60. Singh RK, Mehta S, Jaiswal D, Rai PK, Watal G (2009) Antidiabetic effect of *Ficus bengalensis* aerial roots in experimental animals. *J Ethnopharmacol* 123(1):110–114. <https://doi.org/10.1016/j.jep.2009.02.017>
61. Venkatesh S, Thilagavathi J, Shyam SD (2008) Anti-diabetic activity of flowers of *Hibiscus rosa-sinensis*. *Fitoterapia* 79(2):79–81. <https://doi.org/10.1016/j.fitote.2007.06.015>
62. Al-Snafi A (2018) Chemical constituents, pharmacological effects and therapeutic importance of *Hibiscus rosa-sinensis*. *IOSR J Pharm* 8(7):101–119
63. Panda V, Sonkamble M (2012) Phytochemical constituents and pharmacological activities of *Ipomoea batatas* (Lam). *Int J Res Pharmacol Pharmacother* 2(1):25–34
64. Ogunrinola OO, Fajana OO, Olaitan SN, Adu OB, Akinola MO (2015) Antidiabetic activity of *Ipomoea batatas* leaves extract: effects on hepatic enzymes in alloxan-induced diabetic rats. *Res J Med Plant* 9(5):227–233. <https://doi.org/10.3923/rjmp.2015.227.233>
65. Mishra SB, Vijayakumjar M, Ojha SK, Verma A (2010) Antidiabetic effect of *Jatropha curcas* L. leaves extract in normal and alloxan-induced diabetic rats. *Int J Pharm Sci* 2(1):482–487
66. Saleem M, Tanvir M, Akhtar MF, Iqbal M, Saleem A (2019) Antidiabetic potential of *Mangifera indica* L. cv. Anwar Ratol leaves: medicinal application of food wastes. *Medicina (Kaunas)* 55(7):353. <https://doi.org/10.3390/medicina55070353>
67. Ahn E, Lee J, Jeon YH, Choi SW, Kim E (2017) Anti-diabetic effects of mulberry (*Morus alba* L.) branches and oxyresveratrol in streptozotocin-induced diabetic mice. *Food Sci Biotechnol* 26(6):1693–1702. <https://doi.org/10.1007/s10068-017-0223-y>
68. Chan EW, Lye PY, Wong SK (2016) Phytochemistry, pharmacology, and clinical trials of *Morus alba*. *Chin J Nat Med* 14(1):17–30. <https://doi.org/10.3724/SPJ.1009.2016.00017>
69. Majekodunmi SO, Oyagbemi AA, Umukoro S, Odeku OA (2011) Evaluation of the anti-diabetic properties of *Mucuna pruriens* seed extract. *Asian Pac J Trop Med* 4(8):632–636. [https://doi.org/10.1016/S1995-7645\(11\)60161-2](https://doi.org/10.1016/S1995-7645(11)60161-2)
70. Yadav M, Upadhyay P, Purohit P, Pandey B, Shah H (2017) Phytochemistry and pharmacological activity of *Mucuna pruriens*: a review. *Int J Green Pharm* 11(02):69–77. <https://doi.org/10.22377/jgp.v1i02.916>
71. Gharib E, Montasser Kouhsari S (2019) Study of the antidiabetic activity of *Punica granatum* L. fruits aqueous extract on the alloxan-diabetic wistar rats. *Iran J Pharm Res* 18(1):358–368
72. Ahmed MF, Kazim SM, Ghori SS, Mehjabeen SS, Ahmed SR, Ali SM, Ibrahim M (2010) Antidiabetic activity of *Vinca rosea* extracts in alloxan-induced diabetic rats. *Int J Endocrinol.* <https://doi.org/10.1155/2010/841090>
73. Soren G, Sarita M, Prathyusha T (2016) Antidiabetic activity of *Actinidia deliciosa* fruit in alloxan induced diabetic rats. *Pharma Innov* 5(9):31–34
74. Shirwaikar A, Rajendran K, Kumar CD, Bodla R (2004) Antidiabetic activity of aqueous leaf extract of *Annona squamosa* in streptozotocin—nicotinamide type 2 diabetic rats. *J Ethnopharmacol* 91(1):171–175. <https://doi.org/10.1016/j.jep.2003.12.017>

75. Sabu MC, Smitha K, Kuttan R (2002) Anti-diabetic activity of green tea polyphenols and their role in reducing oxidative stress in experimental diabetes. *J Ethnopharmacol* 83(1–2):109–116. [https://doi.org/10.1016/S0378-8741\(02\)00217-9](https://doi.org/10.1016/S0378-8741(02)00217-9)
76. Nisha M, Vinod B, Christudas S (2018) Evaluation of *Boerhavia erecta* L. for potential antidiabetic and antihyperlipidemic activities in streptozotocin-induced diabetic Wistar rats. *Future J Pharm Sci* 4:150–155. <https://doi.org/10.1016/j.fjps.2017.12.001>
77. Ekowati N, Yuniati N, Hernayanti H, Ratnaningtyas N (2018) Antidiabetic potentials of button mushroom (*Agaricus bisporus*) on alloxan-induced diabetic rats. *Biosaintifika J Biol Biol Educ* 10:655–662. <https://doi.org/10.15294/biosaintifika.v10i3.17126>
78. Ali MA, Wahed MI, Khatune NA, Rahman B, Barman RK, Islam MR (2015) Antidiabetic and antioxidant activities of ethanolic extract of *Seme-carpus anacardium* (Linn.) bark. *BMC Complement Altern Med* 15:138. <https://doi.org/10.1186/s12906-015-0662-z>
79. Fred-JA KA (2009) Antidiabetic activity of *Spondias mombin* extract in NIDDM rats. *Pharm Biol* 47:215–218. <https://doi.org/10.1080/13880200802462493>
80. Sameh S, Al-sayed E, Labib RM, Singab AN (2018) Genus spondias : a phytochemical and pharmacological review. Hindawi. <https://doi.org/10.1155/2018/5382904>
81. Shahwar D, Ullah S, Ahmad M, Ullah S, Ahmad N, Khan M (2012) Hypoglycemic activity of *Ruellia tuberosa* Linn. (Acanthaceae) in normal and alloxan-induced diabetic rabbits. *Iran J Pharm Sci* 7(2):107–115
82. Kumar A, Pathak M, Chaudhary RP, Verma V, Singh L (2022) Pharmacognostical studies and quality control parameters of *Sidarhombifolia*. *IJBPS* 11(2):662–672
83. Nawel M, Dib M, Allali H, Boufelfa T (2011) Hypoglycaemic effect of *Berberis vulgaris* L. in normal and streptozotocin-induced diabetic rats. *Asian Pac J Trop Biomed* 1(6):468–471. [https://doi.org/10.1016/S2221-1691\(11\)60102-0](https://doi.org/10.1016/S2221-1691(11)60102-0)
84. El Barky A, Hussein S, Alm-Eldeen A, Hafez A, Mohamed T (2017) Saponins and their potential role in diabetes mellitus. *Diabetes Manag* 7:148–158
85. Philpott DJ, Butzner JD, Meddings JB (1992) Regulation of intestinal glucose transport. *Can J Physiol Pharmacol* 70(9):1201–1207. <https://doi.org/10.1139/y92-167>
86. Kumari M, Jain S (2012) Tannins: an antinutrient with positive effect to manage diabetes. *Res J Recent Sci* 1(12):70–73
87. Kim MJ, Ryu GR, Chung JS, Sim SS, Min DS, Rhie DJ, Yoon SH, Hahn SJ, Kim MS, Jo YH (2003) Protective effects of epicatechin against the toxic effects of streptozotocin on rat pancreatic islets: in vivo and in vitro. *Pancreas* 26(3):292–299. <https://doi.org/10.1097/00006676-20034000-00014>
88. Kumar D, Ghosh R, Pal B (2013) α -Glucosidase inhibitory terpenoids from *Potentilla fulgens* and their quantitative estimation by validated HPLC method. *J Funct Foods* 5(3):1135–1141. <https://doi.org/10.1016/j.jff.2013.03.010>
89. Mbaze LM, Poumale HM, Wansi JD, Lado JA, Khan SN, Iqbal M, Ngadjui BT, Laatsch H (2007) alpha-Glucosidase inhibitory pentacyclic triterpenes from the stem bark of *Fagara tessmannii* (Rutaceae). *Phytochemistry* 68(5):591595. <https://doi.org/10.1016/j.phytochem.2006.12.015>
90. Eliza J, Daisy P, Ignacimuthu S, Duraiapandian V (2009) Normo-glycemic and hypolipidemic effect of costunolide isolated from *Costus speciosus* (Koen ex. Retz.) Sm. in streptozotocin-induced diabetic rats. *Chem Biol Interact* 179(2–3):329–334. <https://doi.org/10.1016/j.cbi.2008.10.017>
91. Prasad S, Kalra N, Shukla Y (2007) Hepatoprotective effects of lupeol and mango pulp extract of carcinogen induced alteration in Swiss albino mice. *Mol Nutr Food Res* 51(3):352–359. <https://doi.org/10.1002/mnfr.200600113>
92. Panigrahy SK, Bhatt R, Kumar A (2021) Targeting type II diabetes with plant terpenes: the new and promising antidiabetic therapeutics. *Biologia* 76:241–254. <https://doi.org/10.2478/s11756-020-00575-y>
93. Samarghandian S, Borji A, Delkhosh MB, Samini F (2013) Safranal treatment improves hyperglycemia, hyperlipidemia and oxidative stress in streptozotocin-induced diabetic rats. *J Pharm Pharm Sci* 16(2):352–362. <https://doi.org/10.18433/j3zs3q>
94. Naik SR, Barbosa Filho JM, Dhuley JN, Deshmukh V (1991) Probable mechanism of hypoglycemic activity of basic acid, a natural product isolated from *Burmelia sartorum*. *J Ethnopharmacol* 33(1–2):37–44. [https://doi.org/10.1016/0378-8741\(91\)90158-a](https://doi.org/10.1016/0378-8741(91)90158-a)
95. Wang HY, Kan WC, Cheng TJ, Yu SH, Chang LH, Chuu JJ (2014) Differential anti-diabetic effects and mechanism of action of charantin-rich extract of Taiwanese *Momordica charantia* between type 1 and type 2 diabetic mice. *Food Chem Toxicol* 69:347–356. <https://doi.org/10.1016/j.fct.2014.04.008>
96. Tan MJ, Ye JM, Turner N, Hohnen-Behrens C, Ke CQ, Tang CP, Chen T, Weiss HC, Gesing ER, Rowland A, James DE, Ye Y (2008) Antidiabetic activities of triterpenoids isolated from bitter melon associated with activation of the AMPK pathway. *Chem Biol* 5(3):263–273. <https://doi.org/10.1016/j.chembiol.2008.01.013>
97. Tahira S, Hussain F (2014) Antidiabetic evaluation of *Momordica charantia* L. fruit extracts. *West Indian Med J* 63(4):294–299. <https://doi.org/10.7727/wimj.2013.180>
98. Morikawa T, Kishi A, Pongpiriyadacha Y, Matsuda H, Yoshikawa M (2003) Structures of new friedelane-type triterpenes and eudesmane-type sesquiterpene and aldose reductase inhibitors from *Salacia chinensis*. *J Nat Prod* 66(9):1191–1196. <https://doi.org/10.1021/np0301543>
99. Kishi A, Morikawa T, Matsuda H, Yoshikawa M (2003) Structures of new friedelane- and norfriedelane-type triterpenes and polyacylated eudesmane-type sesquiterpene from *Salacia chinensis* Linn. (*S. prionoides* DC. Hippocrateaceae) and radical scavenging activities of principal constituents. *Chem Pharm Bull* 51(9):1051–1055. <https://doi.org/10.1248/cpb.51.105>
100. Zaharudin N, Staerk D, Dragsted LO (2019) Inhibition of α -glucosidase activity by selected edible seaweeds and fucoxanthin. *Food Chem* 270:481–486. <https://doi.org/10.1016/j.foodchem.2018.07.142>
101. Adhikari B (2021) Roles of alkaloids from medicinal plants in the management of diabetes mellitus. *J Chem*. <https://doi.org/10.1155/2021/2691525>
102. Oates PJ (2008) Aldose reductase, still a compelling target for diabetic neuropathy. *Curr Drug Targets* 9(1):14–36. <https://doi.org/10.2174/138945008783431781>
103. Guasch L, Ojeda MJ, Gonzalez-Abuin N, Sala E, Cereto-Massague A, Mulero M, Valls C, Pinent M, Ardevol A, Garcia-Vallve S, Pujadas G (2012) Identification of novel human dipeptidyl peptidase-IV inhibitors of natural origin (part I): virtual screening and activity assays. *PLoS ONE* 7(9):e44971. <https://doi.org/10.1371/journal.pone.0044971>
104. Seino Y, Fukushima M, Yabe D (2010) GIP and GLP-1, the two incretin hormones: similarities and differences. *J Diabetes Investig* 1(1–2):8–23. <https://doi.org/10.1111/j.2040-1124.2010.00022.x>
105. Kim W, Egan JM (2008) The role of incretins in glucose homeostasis and diabetes treatment. *Pharmacol Rev* 60(4):470–512. <https://doi.org/10.1124/pr.108.000604>
106. Tiong SH, Looi CY, Hazni H, Arya A, Paydar M, Wong WF, Cheah SC, Mustafa MR, Awan K (2013) Antidiabetic and antioxidant properties of alkaloids from *Catharanthus roseus* (L.) G. Don. *Molecules* (Basel, Switzerland) 18(8):9770–9784. <https://doi.org/10.3390/molecules18089770>
107. Yen FS, Qin CS, Shi XS, Ying PJ, Le HY, Darmarajan T, Gunasekaran B, Salvamani S (2021) Hypoglycemic effects of plant flavonoids: a review. *Evid Based Complement Alternat Med* 2021:2057333
108. Kapoor R, Kakkar P (2012) Protective role of morin, a flavonoid, against high glucose induced oxidative stress mediated apoptosis in primary rat hepatocytes. *PLoS ONE* 7(8):e41663. <https://doi.org/10.1371/journal.pone.0041663>

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.